

Phytochemical Screening And Antioxidant Property Of *Murraya Koenigii* (Curry Leaf)

¹Kavinilavu G., ²M.Basheera John, ³P.Astalakshmi, ⁴D.Vimala.

¹ Ph.D. Research scholar, ²Assistant Professor, ³ Ph.D. Research scholar, ⁴Ph.D. Research scholar

¹ Department of Zoology,

¹ Queen Mary's College (AUTONOMOUS) (Affiliated to *University of Madras*)
Mylapore, Chennai, Tamilnadu, India.

ABSTRACT - *Murraya koenigii*, also known as curry leaf it is a widely used medicinal plant in India. Family Rutaceae, is highly valued for its medicinal value and distinct aroma. It has several pharmacological properties, such as antioxidant activity and antimicrobial activity etc. The present study focuses on Phytochemical screening and analysis of antioxidant activity from three different extracts of *Murraya koenigii* prepared by the Magnetic stirrer extraction method and to detect the presence of alkaloids, phenols, flavonoids, terpenoids, steroids and other phytochemicals. In DPPH analysis (1,1-diphenyl-2-picrylhydrazyl), antioxidant activity is characterized as a stable free radical with the delocalization centred at 517a nm.

Keywords - *Murraya koenigii*, Curry leaf, Rutaceae, phytochemical, antioxidant.

I. INTRODUCTION

Murrayakoenigii is additionally known as curry leaf, Family Rutaceae, it has Different therapeutic frameworks like Ayurveda and so forth and neighbourhood wellbeing customs are drilled in India, which use a huge number of plants for the treatment of different human and creature illnesses. Those plants utilized for therapeutic design are known as restorative plants. The restorative worth of plants relies upon various dynamic mixtures like sugars, proteins, chemicals, oils, terpenoids, phenolic compounds and so on, which are supportive in working on the life and treatment of illness. Many plants have antimicrobial standards like tannins, rejuvenating ointments also, and other fragrant mixtures. These compounds safeguard the plant from microbial contamination and Deterioration [1,2].

The World Health Organization have been estimated that 80% of the population believes in traditional medicine for their basic health care requirements[3,4,5]. The plant *Murrayakoenigii* (L.) has a place in the family Rutaceae, which is broadly conveyed in practically all pieces of Sri Lanka, India and other South Asian nations. It is generally developed in China and found in tropical and sub-tropical locales, for example, Nigeria, Australia and so forth. The leaves of *Murrayakoenigii* are ordinarily called Curry leaves from antiquated times. Theories leaves are utilized for flavouring and seasoning a large portion of Indian dishes. These leaves contain some restorative qualities which are valuable for people. The *Murrayakoenigii* leaves help to alleviate queasiness, spewing, and acid reflux and help in expanding the stomach related discharges [6,7]. The *Murrayakoenigii* is accepted to have particular restorative properties, for example, anti-oxidant, anti-diabetic, cell reinforcement, antimicrobial, mitigating, anti-carcinogenic and hepato-defensive properties. *Murrayakoenigii* leaves contain the most extravagant wellsprings of mahanimbine, koenigine and mu online which go about as carbazole alkaloids and which helps in a cell reinforcement and anticancer properties [8].

Utilizing plants meds were arranged which are effectively accessible, protected and less expensive than that present-day engineered drugs [9]. The restorative utilization of plants is because of the phytoconstituents present in them. A portion of these synthetics were bioactive and contains biochemical and produce distinct physiological activities in creatures and people. They are ordinarily called phytochemicals or optional metabolites which contain flavonoids, alkaloids, tannins, saponin, phenols, glycoside, steroids and terpenoids and so forth [10].

The presence of significant phytochemicals makes the plant valuable against various illnesses and has a solid of giving valuable medications for human use. Phytochemicals are viewed as auxiliary metabolites delivered at next to no sum as the plant has little requirement for them. They are delivered normally in entire pieces of the plant body; like bark, leaves, stem, root, bloom, and so forth [11]. The amount and nature of phytochemicals present in plant parts might contrast from one section to another [12].

The phytochemical compounds gift in the flowers may acts as a complement for the people by natural antioxidants [13]. Many researchers exhibit that plant life consists of the rich assets of antioxidants found in them. Antioxidants are the sources that allow the human body to shield itself from loose radicals at some stage in the oxidation reactions within the body's metabolism [14]. Antioxidants help in controlling and reducing oxidative damage via dilatory or inhibiting oxidation which is a result of Reactive Oxygen Species (ROS) which concurrently increases the self-lifestyles and satisfaction of the meals [15]. *Murrayakoenigii* have a sturdy antioxidant ability [16].

By keeping this in mind, the present investigation was conducted to study the phytochemical analysis, followed by the antioxidant activity in two different solvent extracts of *Murrayakoenigii*.

II. MATERIALS AND METHODS

CHEMICALS

All chemicals and reagents were used of analytical grade. DPPH, Ethanol, Methanol, Ascorbic Acid, Sulfuric Acid, Millon's reagent, sodium nitrite, Mayer's reagent, Molisch reagent, ferric chloride, ninhydrin, and chloroform were purchased from Scientific Chemicals, Chennai, Tamil Nadu, India.

COLLECTION OF PLANT

The leaf of *Murrayakoenigii* was collected in and around the near vegetative area in Chengalpattu, Tamil Nadu, India.

PLANT EXTRACTION PROCEDURE

Freshly collected leaves samples of *M.koenigii* were washed with fresh water 2–3 times and dried under the shade at room temperature and then blended to powder using an electric blender. Powdered leaves were passed through a 2 mm sieve and stored in a sterile airtight container for further use. Extraction of leaves sample was done in two different extractions: Distilled water 100% and methanol 100%. For this purpose, 10 g of dry powdered leaves were placed in the conical flask of 250 ml capacity and 100 ml of different solvents viz. Water and Methanol were added separately. Flasks were tightly sealed with parafilm and allowed to be shaken vigorously on a magnetic stirrer for 48 h. Extracts were filtered using Whatman No.1 filter paper. The filtrates were then stored in an airtight sample bottle in a refrigerator at -4°C until required.

PHYTOCHEMICAL ANALYSIS OF PLANT EXTRACTS

The presence of phytochemicals is determined based on standard qualitative test procedures [18] of The Aqueous, Ethanol and Methanol Extract of *Murraya koenigii* leaf was and these procedures are as follows:

1. **Test for Acids-Millon's Test:** To 1.0 ml extract, five drops of Millon's reagent were added, heated on a water bath for 5 min. and allowed to cool followed by the addition of 1% sodium nitrite solution. Then observed for the formation of red colour, which indicates the presence of acids.
2. **Test for Alkaloids-Mayer's Test:** To 2.0 ml extract, 2.0 ml concentrated hydrochloric acid followed by a few drops of Mayer's reagent were added and observed for the formation of green colour or white precipitate, which indicates the presence of alkaloids.
3. **Test for Carbohydrates -Molisch's Test:** To 2.0 ml extract, 1.0 ml Molisch's and a few drops of concentrated sulphuric acid were added and observed for the formation of a purple or reddish ring, which indicates the presence of carbohydrates.
4. **Test for Cardiac Glycosides-Ferric Chloride Test:** To 0.5 ml extract, 2.0 ml glacial acetic acid and a few drops of 5% ferric chloride was added. This was under layered with 1.0 ml concentrated sodium hydroxide. Formation of the brown ring at the interface was observed, which indicates the presence of cardiac glycosides.
5. **Test for Flavonoids-Sulphuric Acid Test:** 1.0 ml extract was treated with a few drops of concentrated sulphuric acid and observed for the formation of orange colour.
6. **Test for Glycosides-Sulphuric Acid Test:** To 2.0 ml extract, 1.0 ml glacial acetic acid, 5% ferric chloride and a few drops of concentrated sulphuric acid were added and observed for the formation of greenish-blue colour, which indicates the presence of glycosides.
7. **Test for Phenols-Ferric Chloride Test:** To 1.0 ml extract, 2.0 ml distilled water, followed by a few drops of 10% ferric chloride were added. Formation of blue or green colour was observed, which indicates the presence of phenols.
8. **Test for Proteins-Ninhydrin Test:** To 2.0 ml extract, a few drops of 0.2% ninhydrin were added and heated for 5 min. and observed for the formation of blue colour. This indicates the presence of proteins.
9. **Test for Quinones-Sulphuric Acid Test:** To 1.0 ml extract, 1.0 ml concentrated sodium hydroxide was added and observed for the formation of red colour, which indicates the presence of quinones.
10. **Test for Saponins-Foam Test:** To 1.0 ml extract, 5.0 ml distilled water was added and shaken well in a graduated cylinder for 15 min. lengthwise. The formation of a 1.0 cm layer of foam was observed, which indicates the presence of saponins.
11. **Test for Starch-Iodine Test:** To 2.0 ml extract, few drops of iodine solution was added and observed for the formation of blue-purple colour, which indicates the formation of starch.
12. **Test for Steroids-Salkowski Test:** To 5.0 ml extract, 2.0 ml of chloroform and a few drops of concentrated sulphuric acid were added and observed for the formation of red colour, which indicates the presence of steroids.

13. **Test for Tannins-Ferric Chloride Test:** To 1.0 ml extract, 2.0 ml 5% ferric chloride was added and observed for the formation of dark blue or greenish-black colour, which indicates the presence of tannins.
14. **Test for Terpenoids-Sulphuric Acid Test:** To 0.5 ml extract, 2.0 ml chloroform was added and to this, concentrated sodium hydroxide was added carefully. The formation of red-brown colour at the interface was observed, which indicates the presence of terpenoids.

DPPH FREE RADICAL SCAVENGING ANTIOXIDANT ACTIVITY:

DPPH (1,1-diphenyl-2-picrylhydrazyl) is described as a stable free revolutionary by the goodness of the delocalisation of the extra electron over the particle overall, so the atoms don't dimerise, as would be the situation with most other free extremists. The delocalisation additionally leads to the profound violet tone, described by an assimilation band in ethanol arrangement focused at around 520 nm. At the point when an answer of DPPH is blended in with that of a substance that can give a hydrogen particle, then, at that point, this brings about the decreased structure [18] with the deficiency of this violet tone (even though there would be supposed to be a lingering light yellow tone from the picryl bunch present). Addressing the DPPH revolutionary by $Z\cdot$ and the contributor particle by AH, the essential response is $Z\cdot + AH = ZH + A\cdot$ where ZH is the diminished structure and $A\cdot$ is the free extremist created in this initial step. This last extreme will then, at that point, go through additional responses which control the general stoichiometry, or at least, the number of particles of DPPH decreased (decolourised) by one atom of the reductant. [19]. The Antioxidant activity of aqueous and Methanol leaf extract of *Murrayakoenigi* based on the scavenging property of the 2, 2-diphenyl-1-picrylhydrazyl (DPPH) free stable radical was obtained by following the procedure.

The antioxidant activity of the samples and the standard were assessed based on the radical scavenging effect of the stable 1,1-diphenyl-2-picrylhydrazyl (DPPH) – free radical activity method. The Different concentrations of 100-500 μ g of 1ml Aqueous, Ethanol and Methanol Extract of *Murraya koenigii* leaf is diluted. The working solutions of the test samples were prepared in methanol 0.002% of DPPH, 3.7 ml of absolute methanol in all test tubes and 3.8 ml of absolute methanol was added to blank. Add 100 μ l of Ascorbic Acid to the tube marked as standard and 100 μ l of respective samples to all other tubes marked as tests. 200 μ l of DPPH reagent was added to all the test tubes including blank. Incubate all test tubes at room temperature in dark conditions for 30 minutes. The absorbance of all samples was read at 517nm in a spectrophotometer. For tremendous control, Ascorbic acid becomes used. The DPPH loose radical scavenging assets became the optical density was recorded and % inhibition was calculated using the formula given below.

CALCULATION

$$\% \text{ of Antioxidant activity} = \frac{(\text{Absorbance at blank}) - (\text{Absorbance at test})}{(\text{Absorbance at blank})} \times 100$$

STATISTICAL ANALYSIS

Data obtained from the experimental Analysis was expressed in the pattern as Mean and Standard Deviation.

III. RESULT & DISCUSSION

TABLE: 1 - Phytochemical Analysis of *Murraya Koenigii*

S.no	Test	Method	Aqueous	Ethanol	Methanol
1	Amino Acids	Million's Test	-	-	-
2	Alkaloids	Mayer's Test	+	+	+
3	Carbohydrates	Molisch's Test	+	+	+
4	Cardiac Glycosides	Ferric Chloride Test	-	+	+
5	Flavonoids	Sulphuric Acid Test	-	-	+
6	Glycosides	Sulphuric Acid Test	+	+	+
7	Phenols	Ferric Chlorid e Test	+	+	+
8	Proteins	Ninhydrin Test	+	+	+
9	Quinones	Sulphuric Acid Test	+	-	-
10	Saponins	Foam Test	-	+	+
11	Starch	Iodine Test	-	-	-
12	Steroids	Salkowski Test	+	+	+

13	Tannins	Ferric Chloride Test	-	+	+
14	Terpenoids	Sulphuric Acid Test	+	+	+

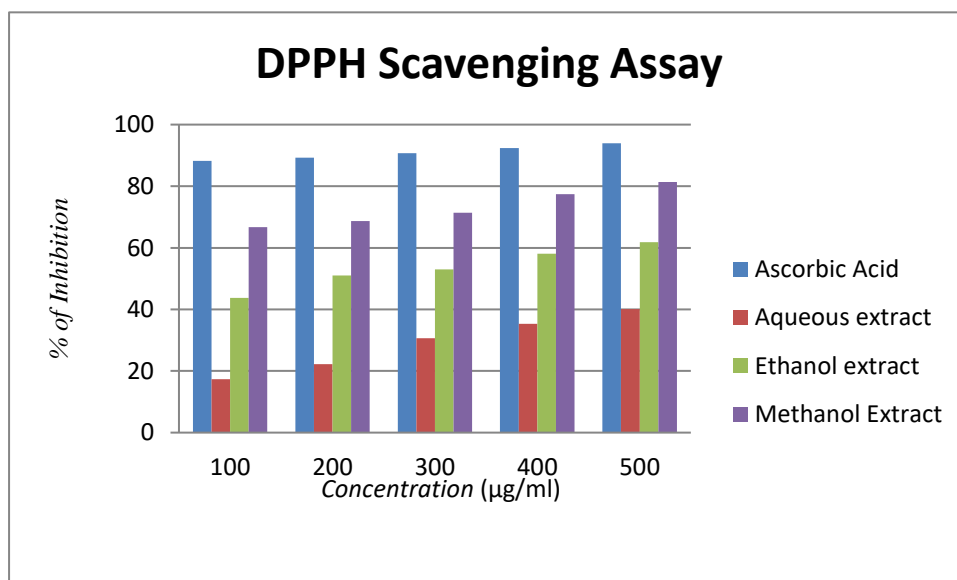
+ = Presence; - = Absence

TABLE: 2- Antioxidant activity of *MurrayaKoenigii*.

DPPH % of inhibition of samples at different concentrations				
Concentrations of the sample (µg/ml)	Standard Ascorbic Acid	Aqueous extract	Ethanol extract	Methanol extract
100	88.21±0.29	31.64±0.29	51.01±0.50	68.68±0.50
200	89.22±0.29	35.35±0.50	53.03±0.50	71.38±0.77
300	90.74±0.58	40.06±0.29	58.08±0.50	77.44±1.27
400	92.42±0.50	43.77±0.29	61.78±0.50	81.31±0.50
500	93.93±0.50	45.45±0.50	66.66±0.77	84.17±0.29

Mean ± Standard deviation (SD) for analysis in three replicates

Figure 1. Total antioxidant activity of Aqueous, Ethanol and methanol extract of *Murrayakoenigii* and Standard Ascorbic acid.



The healing fee of medicinal plant life lies inside the various chemical compounds processed in them. The bioactive compounds of plant extract are attributed to phytochemical compounds of flora. If the flora wealthy in tannin compound they have got highly effective in controlling the bacteria, because of this character they permit to react with the proteins to form solid water-soluble compounds consequently, it kills the bacterial by way without delay damaging its mobile membrane [20]. The DPPH radical scavenging assay has been broadly used to analyze the perspective of the compounds which include free radical scavengers of the hydrogen donors and used to have a look at the antioxidant interest of plant extract. Phenolic compounds present in plant life act as antioxidants or unfasted radical scavengers because of their OH organizations, which are committed directly to the antioxidant action [21].

Qualitative Analysis of phytochemical screening of the three different extracts of *Murraya koenigii* leaves covered the presence of various chemical substance gatherings like Alkaloids, Carbohydrates, Glycosides, Phenol, Proteins, Quinones, steroids and terpenoids in Aqueous Extracts. Alkaloids, Carbohydrates, Cardiac Glycosides, Glycosides, Phenol, Proteins, saponins, Steroids, Tannins and Terpenoids in Ethanol Extracts Furthermore, Alkaloids, Carbohydrates, Cardiac Glycosides, Flavonoids, Glycosides, Phenol, Proteins, saponins, Steroids, Tannins and Terpenoids in Methanol Extracts [Table 1].

The Antioxidant activity of *MurrayaKoenigii* using the DPPH radical scavenging assay is based on the ability of antioxidants, to decolourize DPPH. The DPPH revolutionary contains an odd electron, which is liable for the absorbance at 517 nm. Cell reinforcements give an electron to DPPH and decolourize it, which can be quantitatively estimated from the changes in absorbance. Each of the evaluated concentrates of *Murrayakoenigii* had the option to diminish the stable, purple coloured extremist DPPH to the yellow-shaded DPPH.

The leaf extract of *Murraya koenigii* and standard ascorbic acid revealed the highest percentage of inhibitory activity in 500 µg/ml (93.93±0.50%) in ascorbic acid, (84.17±0.29%) in Methanol, (66.66±0.77%) in Ethanol and (45.45±0.50%) in Aqueous. The leaf extract of *Murraya koenigii* and standard ascorbic acid revealed the least percentage of inhibitory activity in 100 µg/ml (31.64±0.29%) in Aqueous, (51.01±0.50%) in Ethanol, (68.68±0.50%) in Methanol, and (88.21±0.29%) in ascorbic acid. [Table 2 & Figure 1] from the above study, *Murraya koenigii* exhibits good Results.

IV. CONCLUSIONS

In this current above study, it very well may be pronounced that the phytochemical screening and anti-oxidant DPPH activity of Aqueous and Methanol extract of *Murrayakoenigii* leaf extract have good properties of Phytochemical and Antioxidant. The methanol Extract has very good properties than the Aqueous Extract. *Murrayakoenigii* contains the functional groups that can be implemented in pharmaceutical industries to develop drugs to cure many diseases.

ACKNOWLEDGEMENTS

We thank the anonymous referees for their useful suggestions.

REFERENCES

1. Cowan, M.M. 1999. Plant products as antimicrobial agents. *Clin Microb Rev.*12: 564–82p.
<https://journals.asm.org/doi/abs/10.1128/cmr.12.4.564>
2. Rashmi, Jyoti Bisht, Naveen G., 2016. Phytochemical Analysis and Antibacterial Activity of Different Leaf Extracts of *Murrayakoenigii*. *journalsPup* Vol. 2: Issue 2., 1-5
https://www.academia.edu/download/51803368/antibacterial_activity_of_M._koenigii.pdf
3. Sindhu RK, Arora S.. 2012. Phytochemical and Pharmacognostical studies on *Murrayakoenigii* L. spreng roots. *Drug Invention Today.*; 4:325-336.
<https://search.ebscohost.com/login.aspx?direct=true&profile=ehost&scope=site&authtype=crawler&jrnl=09757619&AN=120376583&h=%2FPflz%2FITr5TGEHWY688vNwaENqh%2BV6LgYZQujAcsVdEq4fotL%2FUBr6hpAkhuvTZZ6WjAmha45GL%2BVonaBPo%2Fdw%3D%3D&crI=c>
4. Kurain JC: In *Plants That Heal*, Oriental Watchman Publishing House, Pune, India. 1995; 296.
5. Aju BY, Rajalakshmi R and Mini S. (2017) Evaluation of the antioxidant activity of *Murrayakoenigii* (L.) Spreng using different *in vitro* methods 6(4): 939-942
6. Chevallier, A. (1996). *The Encyclopedia of Medicinal Plants*. London/New York/Stuttgart/Moscow: Dorling Kindersley, 366. <https://www.cefracor.org/sites/www.cefracor.org/files/webform/documents/offre-complete/fichier/pdf-the-encyclopedia-of-medicinal-plants-andrew-chevallier-pdf-download-free-book-e34b9db.pdf>
7. Deepika, T., C.M. Noorjahan (2019) Phytochemical Screening and Antioxidant Property of Aqueous *Murrayakoenigii* (Curry Leaf) Extract. *IJRAT*, Vol.7, No.2
[https://scholar.google.com/scholar?hl=en&as_sdt=0%2C5&q=7.%09Deepika%2CT.%2C+C.M.+Noorjahan%282019%29+Phytochemical+Screening+and+Antioxidant+Property+of+Aqueous+Murraya+Koenigii+%28Curry+Leaf%29+Extract.+IJRAT%2C+Vol.7%2C+No.2+&btnG=#:~:text=%5BCITATION%5D-.Phytochemical%20Screening%20and%20Antioxidant%20Property%20of%20Aqueous%20Murraya%20Koenigii%20\(Curry%20Leaf\)%20Extract.-T%20Deepika%2C%20CM](https://scholar.google.com/scholar?hl=en&as_sdt=0%2C5&q=7.%09Deepika%2CT.%2C+C.M.+Noorjahan%282019%29+Phytochemical+Screening+and+Antioxidant+Property+of+Aqueous+Murraya+Koenigii+%28Curry+Leaf%29+Extract.+IJRAT%2C+Vol.7%2C+No.2+&btnG=#:~:text=%5BCITATION%5D-.Phytochemical%20Screening%20and%20Antioxidant%20Property%20of%20Aqueous%20Murraya%20Koenigii%20(Curry%20Leaf)%20Extract.-T%20Deepika%2C%20CM)
8. Kirupa, SLS.; Kariitha, R. (2015): Anti-oxidant enhancing property of curry leaf powder *M. koenigii* in type II Diabetes mellitus. *Intl. Journal of Pharmacy and Biosciences*, 6(1), pp: 507-514.
https://scholar.google.com/scholar?hl=en&as_sdt=0%2C5&q=8.%09Kirupa%2C+SLS.%3B+Kariitha%2C+R.+%282015%29%3A+Anti-oxidant+enhancing+property+of+curry+leaf+powder+M.+koenigii+in+type+II+Diabetes+mellitus.+Intl.+Journal+of+Pharmacy+and+Biosciences%2C+6%281%29%2C+pp%3A+507-514.&btnG=#:~:text=Anti%2Doxidant%20enhancing%20property%20of%20curry%20leaf%20powder%20Murraya%20koenigii%20in%20type%20II%20diabetes%20mellitus
9. Kaur, G.; Daftardar, S.; Barve.; KH. (2014): Modifying anti-inflammatory effect of Diclofenac with *Murrayakoenigii*: Recent patent on inflammation and allergy. *Drug Discovery*, 8(1), pp: 77-81.
<https://www.ingentaconnect.com/content/ben/iad/2014/00000008/00000001/art00010>
10. Summon, S.; More, PK.; Sandhya, MM. (2014): Curry leaves (*Murrayakoenigii* Linn Spreng) a miracle plant. *Indian Journal of Science Research*, 4(1), pp: 46-52.
https://www.academia.edu/download/33894505/602447219Microsoft_Word_-_paper_3.pdf
11. Tiwari P.; kumar, B.; Kaur M.; Kaur, G. and Kaur, H. 2011. Phytochemical screening and extraction: A review. *Int.Pharm.Sci.*1(1):98–106p
https://scholar.google.com/scholar?hl=en&as_sdt=0%2C5&q=11.%09Tiwari+P.%3B+kumar%2C+B.%3B+Kaur+M.%3B+Kaur%2C+G.+and+Kaur%2C+H.+2011.+Phytochemical+screening+and+extraction%3A+A+review.+Int+Pharm+Sci.+1%281%29%29

3A+98%E2%80%93106p&btnG=#:~:text=%5BCITATION%5D-

- .Phytochemical%20screening%20and%20extraction%20a%20review,M%20Kaur%2C%20G%20Kaur%2C%20H%20Kaur%2C%20A%20D%20Internationale%20pharmaceutica%20scientia%2C%202011,-Save%20Cite%20Cited
12. Lahlou, M. 2004. Methods to study the phytochemistry and bioactivity of essential oils. *Phytother Res.* 18: 435–45p. <https://onlinelibrary.wiley.com/doi/abs/10.1002/ptr.1465>
 13. Boots, AW., Haenen, GR., Bast, A. (2008). Health effects of quercetin: From antioxidant to nutraceutical. *Eur. J. Pharmacol*, 585, pp:325–337. <https://www.sciencedirect.com/science/article/pii/S0014299908002884>
 14. Farasat, M.; Khavari-Nejad, RA.; Nabavi, SMB.; Namjooyan, F. (2014): Antioxidant activity, Total Phenolics and Flavonoid contents of some edible Green Seaweeds from Northern Coasts of the Persian Gulf. *Iran Journal of Pharma Res.*, 13(1), pp: 163-170. <https://www.ncbi.nlm.nih.gov/pmc/articles/PMC3985267/>
 15. Ruan, ZP.; Zhang, LL.; Lin, YM. (2008): Evaluation of the antioxidant activity of *Syzygiumcuminileaves*. *Molecules*, 13, pp: 2545–2556. <https://www.mdpi.com/6618>
 16. Aju, BY.; Rajalakshmi, R.; Mini, S. (2017): Evaluation of the antioxidant activity of *Murrayakoenigii(L.) Spreng* using different *in-vitro* methods. *Journal of Pharmacognosy and Phytochemistry*, 6(4), pp: 939-942.
 17. Harborne, JB. (1973): *Phytochemical methods: A guide to modern techniques of plant analysis.* Chapman and Hall Ltd. London, pp: 49-279. [https://scholar.google.com/scholar?lookup=0&q=19.%09Harborne,+JB.+\(1973\):+Phytochemical+methods:+A+guide+to+modern+techniques+of+plant+analysis.+Chapman+and+Hall+Ltd.+London,+pp:+49-279.+&hl=en&as_sdt=0,5#:~:text=JB%20Herborne%C2%A0D%20A%20guide%20to%20modern%20techniques%20of%20pant%20analysis%2C%201973](https://scholar.google.com/scholar?lookup=0&q=19.%09Harborne,+JB.+(1973):+Phytochemical+methods:+A+guide+to+modern+techniques+of+plant+analysis.+Chapman+and+Hall+Ltd.+London,+pp:+49-279.+&hl=en&as_sdt=0,5#:~:text=JB%20Herborne%C2%A0D%20A%20guide%20to%20modern%20techniques%20of%20pant%20analysis%2C%201973)
 18. Blois, MS. (1958). Antioxidant determination by the use of a stable free radical. *Nature*, 181, pp: 1199-1200. [https://www.scirp.org/\(S\(i43dyn45teexjx455qlt3d2q\)\)/reference/ReferencesPapers.aspx?ReferenceID=1148740](https://www.scirp.org/(S(i43dyn45teexjx455qlt3d2q))/reference/ReferencesPapers.aspx?ReferenceID=1148740)
 19. Molyneux, P., Songklanakarin *J. sci. technol*, 2004 - The use of the stable free radical diphenylpicrylhydrazyl (DPPH) for estimating antioxidant activity 26(2): 211-219. <https://www.researchgate.net/file.PostFileLoader.html?id=503cd237e4f0761a4b000020&assetKey=AS%3A273786924929035%401442287297816>
 20. Mohamed Sham Shihabudeen, H.; Hansi Priscilla, D.; Kavitha, T. (2010): Antimicrobial activity and phytochemical analysis of selected Indian folk medicinal plants. *Int. J. of Pharma Sci Res.*, 1(10), pp: 430-434. <https://citeseerx.ist.psu.edu/viewdoc/download?doi=10.1.1.702.9581&rep=rep1&type=pdf>
 21. DejanOrcic, Z.; Neda Mimica-Dukic, M.; Marina Franciskovic, M.; Slobodan Petrovic, S.; EmilijaJovin, D. (2011): Antioxidant activity relationship of phenolic compounds in *Hypericum perforating L.* *Chem. Cent. J.*, 5:34. <https://link.springer.com/article/10.1186/1752-153X-5-34>