

Assessment of the prevalence of periodontal pathogens in the plaque and gingiva of patients with fixed and removable orthodontic appliance

Type of manuscript: Research article

Running Title: Periodontal pathogens in fixed and removable orthodontic appliance

Kadambari Sriram, Dr.Sri Rengalakshmi.M

Undergraduate Student, Senior Lecturer
Department of Orthodontics
Saveetha Dental College, Saveetha University, Chennai, India.

Corresponding Author

Dr.Sri Rengalakshmi.M
Senior Lecturer
Department of Orthodontics
Saveetha Dental College
Saveetha University
162, Poonamalle High Road
Chennai 600077
Tamil Nadu, India

ABSTRACT:

AIM:

To assess the prevalence of periodontal pathogens in patients with fixed and removable orthodontic appliance.

INTRODUCTION:

Fixed and removable orthodontic appliances may complicate an optimal oral hygiene, and this may result in accumulation of dental plaque and gingival inflammation. Evidence indicates the gram-negative obligate anaerobe *Porphyromonas gingivalis* and *A.actinomycescomitans* as putative periodontal pathogens in subgingival dental plaque. *P. gingivalis* plays an important role in the onset and progression of periodontal diseases, and it is implicated as an indicator of periodontal disease.

The aim of this study was to evaluate the occurrence of periodontal pathogens in patients with clinical manifestation of plaque-associated gingivitis treated with fixed and removable orthodontic appliances.

MATERIALS AND METHODS:

The study was done using convenience sampling of 10 patients undergoing orthodontic treatment from the Orthodontics Department of Saveetha Dental College, Chennai, out of whom 5 were undergoing fixed orthodontic treatment and 5 were wearing removable orthodontic appliances. The study compared the growth of two periodontal pathogens, *Porphyromonas gingivalis* and *Actinobacillus actinomycescomitans* in the plaque deposits collected during the orthodontic treatment. Subgingival plaque samples were collected by inserting a sterile dental curette into the bottom of the gingival crevice during the clinical examination. The samples were collected from the labial-medial and labial-distal surfaces of teeth 31, 32, 41, and 42. The samples from each tooth were pooled into an Eppendorf tube containing 1 mL of sterile saline and stored immediately at -70 degree Centigrade. Analysis was carried out by Real-time PCR.

RESULT:

Quantification of organisms, *Porphyromonas gingivalis* and *Actinobacillus actinomycescomitans*, have been done for the samples F1,F2,F3,F4,F5 and RA1, RA2, RA3, RA4, RA5 by quantitative PCR (qPCR). It has been found that sample F2 and RA4 has a maximum count of 4178.61943 and 565.56 per ml respectively of *P.gingivalis*.

It has also been found that sample F2 and RA5 has a maximum count of 3821.7132 and 231.59 per ml of *A.actinomycescomitans* respectively.

CONCLUSION:

To create awareness about the occurrence of plaque-associated gingivitis due to periodontal pathogens in patients treated with fixed and removable appliances.

KEYWORDS: Fixed orthodontic treatment, Removable appliance, Periodontal examination, *Porphyromonas gingivalis*, *Actinobacillus actinomycescomitans*, Real-time PCR

INTRODUCTION:

Fixed orthodontic therapy is an effective and common method for treating malocclusions in contemporary orthodontics. Fixed appliances such as brackets, bands, or fixed retentions may complicate an optimal oral hygiene, and this may result in accumulation of dental plaque and gingival inflammation^{1,2}.

One of the common side effects during orthodontic therapy are gingivitis and periodontitis. It is still unclear whether the periodontal changes in orthodontic therapy could be reversible after the removal of appliances. Most studies indicated that gingival changes were only temporary and could be reversible³, while a few of researches reported a significantly clinical attachment loss during orthodontic therapy⁴⁻⁶. A prospective study discovered that orthodontic accessories had a negative impact on periodontal parameters, moreover these changes were only partially reversible post therapy⁵. It has been recognized that anaerobic microorganisms in the subgingival plaque are the key etiologic factors in the initiation and progression of gingivitis and periodontitis⁷. Recent advancements in the periodontal research field supported the theory that periodontal diseases are resulted from a rupture of the dynamic balance between the relative abundance of periodontopathogens and host defence system⁸. Positive associations between periodontal diseases and several pathogens have been reported, including *A. actinomycetemcomitans* and *P. gingivalis*⁹⁻¹¹.

Ample evidence indicates that gram-negative obligate anaerobe *Porphyromonas gingivalis* plays an important role in the onset and progression of periodontal diseases, and it is implicated as an indicator of periodontal disease¹²⁻¹⁶. Gingival changes and oral bacteria in plaque have been studied during orthodontic treatment in adolescents and young adults¹⁷⁻¹⁹. Previous researches revealed that *P. gingivalis* has been categorized as the “red complex” species, which are related to the severity of periodontitis, while *A. actinomycetemcomitans* is categorized as secondary harmful species involved in periodontitis²⁰⁻²³. Additionally, current evidence suggested orthodontic appliances could alter the equilibrium of the microorganism ecosystem, and increased the potential for pathogenicity within the microbial ecosystem^{24, 25}. It is crucial to understand the composition and changes of periodontopathogens during orthodontic therapy in order to avoid potentially irreversible injuries caused by orthodontic appliances.

Socransky et al. showed that the presence of fixed orthodontic appliances encouraged the growth of periodontopathic bacteria species such as *Porphyromonas gingivalis* and *Actinobacillus actinomycetemcomitans*. Nevertheless, the use of alternative removable orthodontic appliances may allow patients to maintain an adequate oral hygiene and reduce the risk for such negative dental and periodontal complication²⁷⁻³⁰.

There is little information on the microbiological evaluation of the subgingival pathogenic microflora via real-time PCR analyses which may be a suitable method to estimate the risk for periodontal disease³¹⁻³³.

In this study, we quantify subgingival pathogens of *A. actinomycetemcomitans*, *P. gingivalis* with the real-time PCR and tested the clinical parameters in adolescents during orthodontic treatment to assess whether the microbial and periodontal parameters are different between removable and fixed orthodontic appliances.

MATERIALS AND METHODS:

Polymerase Chain Reaction (PCR)

The 3 steps of PCR are repeated for about 30 to 40 times in an automated thermal cycler, which heat and colds the reaction mixture in the tube in a very short time. This result in exponential increase accumulation of the specific DNA fragments.

Quantification methods:

Relative quantification

Relative quantification was used in this study as it was needed to compare changes in multiple samples which varied in quality and quantity.

Relative ratio = concentration of target / concentration of reference

Primer Details:

Organisms Name	Primer Sequences (5'-3')
Porphyromonas gingivalis	FP: GCGTAGGTTGTTCCGGTAAGT RP: CATACTGCGACTGACACTGAA
Actinobacillus actinomycetemcomitans	FP : GCCCGAAGCTAACGTGATAA RP: CTTCGGATGTCAAGAGTAGGTAAG

Procedure

1. Take out the samples and all the stocks (from -20°C) needed for Real Time PCR, keep them on ice and allow them to thaw.
2. PCR reaction are set as shown in the Table 1
3. If there are many samples to be analyzed, it is advisable to make cocktail for as many numbers of samples. Cocktail can be prepared with one or two extra volumes so that shortage of reaction mix can be avoided due to pipetting error.
4. The SYB Green I Master mix is light sensitive.
5. The mixture of SYB Green I Master mix, water and primer was mixed, vortexed and spun for few seconds.
6. This reaction mix was loaded onto the Real time PCR plate.
7. Then the respective sample was added to the plate.

8. The plate is spun for few seconds.
9. NTC is mandatory in real time experiments.
10. qPCR was set according to the program given below (Table.2)

Components	Volume (μ l)
Sample	5.0
SYB Green I Master mix (2X)	10
Sterilized water	5
Total Volume	20

Table 1: PCR reaction Mixture

qPCR PROGRAM

Step	Temperature	Temperature	No. of cycles
Initial Denaturation	95°C	5 min	1
Denaturation	95°C	20 sec	40
Annealing	58°C	20 sec	
Extension	72°C	20 sec	
MC	95°C- 60°C - 95°C	1 min	1
Cooling	40°C	1min	1

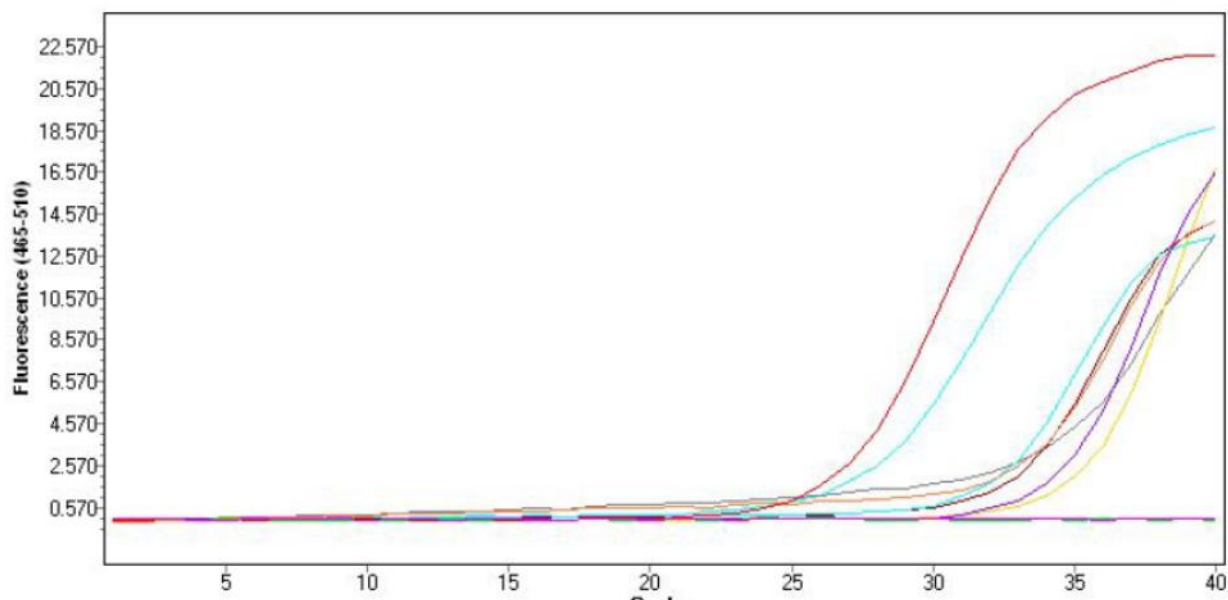
Table 2: qPCR cyclic conditions**RESULTS:****Porphyromonas gingivalis**

FIG:1 Porphyromonas gingivalis

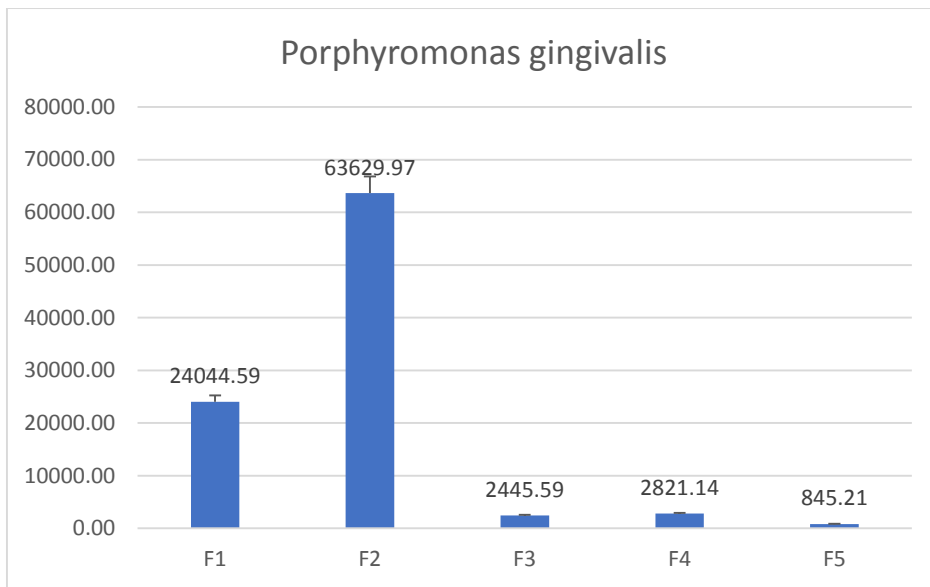


FIG 2: Porphyromonas gingivalis count per mL solution in patients undergoing fixed orthodontic treatment

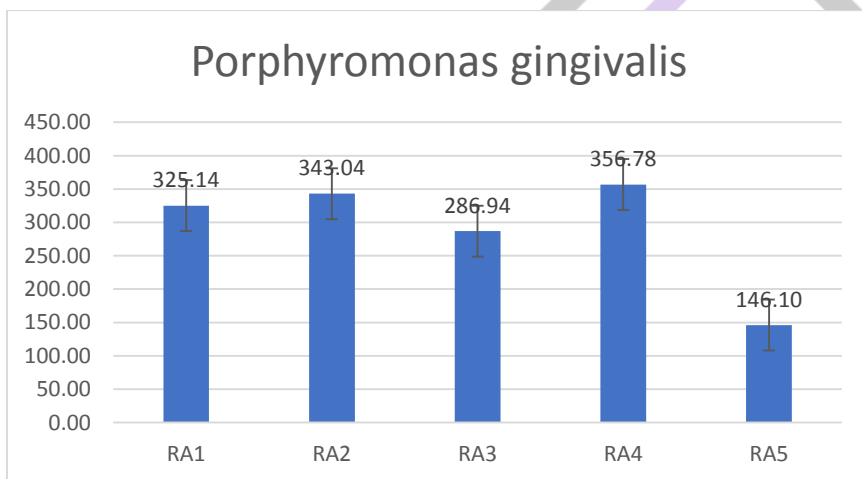


FIG 3: Porphyromonas gingivalis count per mL solution in patients undergoing removable orthodontic treatment.

Actinobacillus actinomycetemcomitans

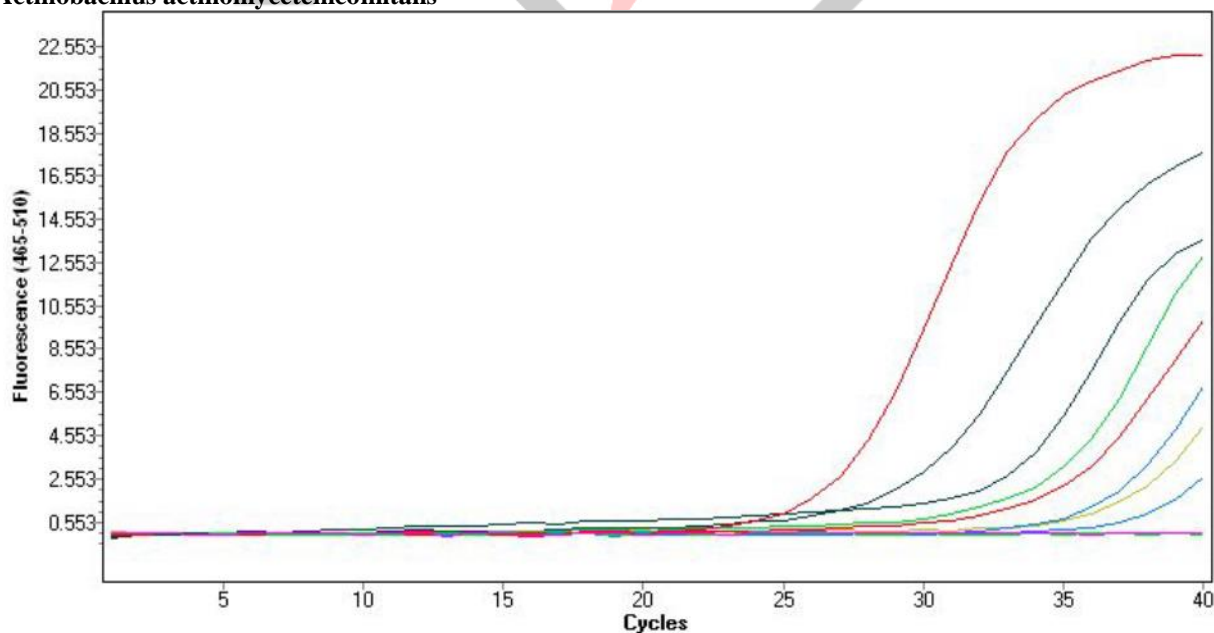


FIG 4: Actinobacillus actinomycetemcomitans

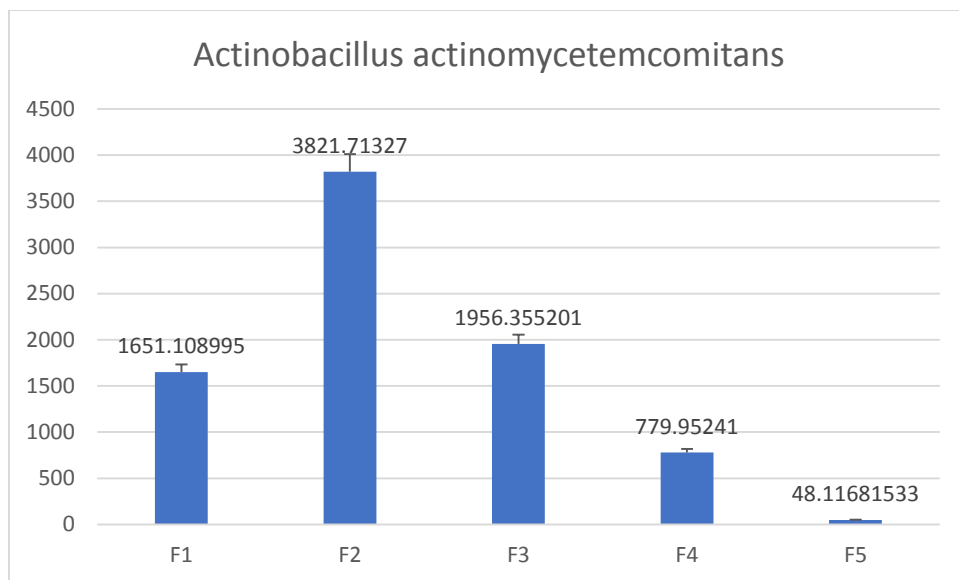


FIG 5: Actinobacillus actinomycetemcomitans Count per ml of the solution in patients undergoing fixed orthodontic treatment

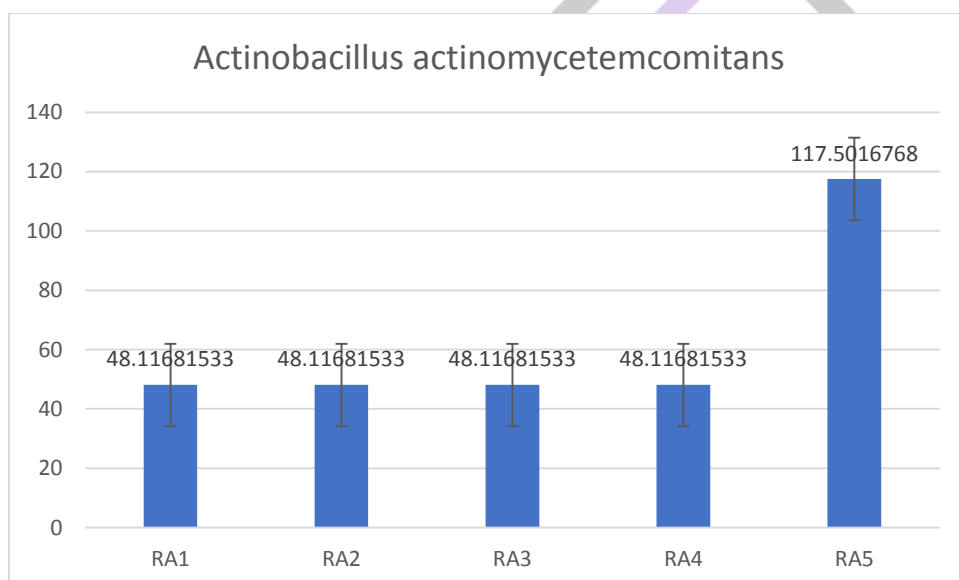


FIG 6: Actinobacillus actinomycetemcomitans Count per ml of the solution in patients undergoing removable orthodontic treatment

TABLE 3: Count per mL of P. gingivalis and A. actinomycetemcomitans in Fixed appliance patients

Samples	F1	F2	F3	F4	F5
Porphyromonas gingivalis	24044.59	63629.97	2445.59	2821.14	845.21
Actinobacillus actinomycetemcomitans	1651.108995	3821.71327	1956.355201	779.95241	48.11681533

TABLE 4: Count per mL of P. gingivalis and A. actinomycetemcomitans in Removable appliance patients

Samples	RA1	RA2	RA3	RA4	RA5
Porphyromonas gingivalis	325.14	343.04	286.94	356.78	146.10
Actinobacillus actinomycetemcomitans	48.11681533	48.11681533	48.11681533	48.11681533	117.5016768

Quantification of organisms Porphyromonas gingivalis and Actinobacillus actinomycetemcomitans has been done for the samples F1,F2,F3,F4,F5 and RA1,RA2,RA3,RA4,RA5 by quantitative PCR (qPCR).

Porphyromonas gingivalis

It has been found that sample F2 and RA4 has a maximum count of 4178.61943 and 565.56 per ml respectively. (Fig 1,2 and 3)

Actinobacillus actinomycetemcomitans

It has been found that sample F2 and RA5 has a maximum count of 3821.7132 and 231.59 per ml respectively. (Fig 4,5 and 6)

DISCUSSION:

The role of microorganisms in the aetiopathogenesis of individual periodontal diseases has been discussed in the literature for many years⁵³. This study compared the prevalence of two periodontal pathogens, *Porphyromonas gingivalis* and *Aggregatibacter actinomycetemcomitans* in patients undergoing fixed and removable orthodontic treatment. *P.gingivalis* is an obligate anaerobe while *A.actinomycetemcomitans* is a facultative anaerobe with both pathogens playing a significant role in the onset of periodontitis. The results of our study was done using Real-time PCR analysis on the supragingival plaque samples of the study participants. It was found in our study that the growth of the periodontal pathogens *P.gingivalis* and *A.actinomycetemcomitans* was greater in the samples of patients undergoing fixed orthodontic treatment as compared to patients undergoing removable orthodontic treatment. This is in accordance with previous studies which show *Porphyromonas gingivalis* and *A.actinomycetemcomitans* have the capacity to penetrate into the buccal epithelial cells in the oral cavity. The released buccal epithelial cells can participate in the transmission of periodontal pathogens among individual localities in the same individual or among several individuals^{34,50}. Leung et al. reported that the capacity of bacteria, especially

Actinobacillus actinomycetemcomitans to invade buccal epithelial cells increased after fixing the orthodontic appliance¹⁸; a probable reason being physical damage of cells by individual components of the orthodontic appliance. It was further found in our study that there was greater prevalence of *P.gingivalis* than *A.actinomycetemcomitans* in both the fixed appliance as well as removable appliance patients. This is consistent with a study by Baehni et al, according to which *P.gingivalis* was associated with destructive periodontitis³⁵.

According to previous studies, authors have demonstrated that the occurrence of bacteria in supragingival and subgingival plaque is similar and supragingival plaque can thus serve as a reservoir for bacteria that can subsequently invade the subgingival spaces³⁶. The assessment of occurrence of the individual pathogens confirmed the presence of *Porphyromonas gingivalis* only in the periodontitis patients. Zadeh et al.suggested that destruction of the periodontium induced by *Actinobacillus actinomycetemcomitans* (*A.a.*) is caused by the interaction between this pathogen and immune response of the host³⁷. Its presence may be considered a risk factor for the development of periodontopathies. Okada et al. analysed the occurrence of *Actinobacillus actinomycetemcomitans* and *Porphyromonas gingivalis* in a study with 104 participants concluding that a higher frequency of pathogens studied in the examined set was related to the presence of the fixed orthodontic appliance³⁸. The increased pathogenicity of the dental plaque and the concomitant periodontal changes during orthodontic treatment have been described by several authors (Petti et al., 1997; Naranjo et al., 2006; van Gastel et al., 2008). Several studies showed that the frequencies and counts of periodontopathogens significantly decreased after appliances removal⁴⁰⁻⁴⁵. Kim et al, stated that *P.gingivalis* dramatically decreased immediately after appliances removal, but *A.actinomycetemcomitans* remained unchanged⁴⁵. This is due to the fact that the gingival enlargement induced by orthodontic appliances might provide a favorable environment for the colonization and maturation of anaerobic bacteria, and favors a qualitative shift from a predominance of aerobic bacteria to more putative anaerobic periodontal pathogens^{19,46}. Therefore, removing the orthodontic appliances eliminates their plaque-retentive effect, which might make practicing good oral hygiene easier¹⁹. Additionally, this discrepancy of nonsignificant difference in the count of *A. actinomycetemcomitans* as opposed to significant differences in the count of *P. gingivalis* may be ascribed to the fact that *A. actinomycetemcomitans* is a gram-negative facultative anaerobe, while *P. gingivalis* is an obligate anaerobes, the growth of latter are easily to be stimulated by the ecologic environment induced by gingival enlargement.

Only a few studies have dealt with the effect of a fixed orthodontic appliance on the oral bacterial flora. Lee et al. reported that there were no differences in the frequency of occurrence of *Actinobacillus actinomycetemcomitans* and *Porphyromonas gingivalis* compared to the control group of adult patients with gingivitis and without the orthodontic appliance⁴⁷. Conversely, Paolantonio et al. proved that *Actinobacillus actinomycetemcomitans* colonised subgingival plaque only on teeth with an attached fixed orthodontic appliance¹⁷. Armitage stated that periodontally healthy subjects carry putative periodontal pathogens as part of the normal oral plaque because these bacteria are detected at low numbers in periodontally healthy individuals^{48,49}. These microorganisms may be opportunistic pathogens, the levels of which increase to a critical threshold to induce periodontal tissues destruction. This implies that an increased number of periodotopathogens, rather than frequency, might be an important determinant in the development of periodontal inflammation. Thus, it is reasonable to focus on the numbers of periodontopathogens to prevent periodontal diseases in orthodontic patients.

CONCLUSION:

The growth of *P. gingivalis* and *A.actinomycetemcomitans* were found to be significantly more abundant in fixed appliances when compared to removable appliances. This study quantified the subgingival pathogens as there is no extensive quantification done in previous studies.

In present times the role of periodontal pathogens in the origin and development of general diseases, such as cardiovascular diseases, cerebral vascular diseases, and low birth weight in infants, has been considered more and more frequently. When performing various interventions in the oral cavity (e.g. in professional hygienic treatment), it is necessary to keep in mind that periodontal pathogens

may penetrate into the patient's system. Prevention of any such diseases can be brought about by maintenance of proper oral hygiene in patients with fixed orthodontic appliance as well as suitable antimicrobial measures.

ACKNOWLEDGEMENTS:

We are grateful to the patients for their contribution to the work. We also want to thank Biozone Research Technologies, Chennai, for their assistance in bacteria culture. This work was supported by Saveetha dental College, Chennai.

REFERENCES:

1. Wennstrom JL. Mucogingival considerations in orthodontic treatment. *Semin Orthod.* 1996;2:46–54.
2. Bollen AM, Cunha-Cruz J, Bakko DW, Huang GJ, Hujoel PP. The effects of orthodontic therapy on periodontal health: a systematic review of controlled evidence. *J Am Dent Assoc.* 2008;139:413–422.
3. Abiko Y, Sato T, Mayanagi G, Takahashi N. Profiling of subgingival plaque biofilm microflora from periodontally healthy subjects and from subjects with periodontitis using quantitative real-time PCR. *Journal of periodontal research*, 2010; 45(3):389±95. doi: 10.1111/j.1600-0765.2009.01250.x PMID: 20337892
4. Aass AM, Albandar J, Aasenden R, Tollefsen T, Gjermo P. Variation in prevalence of radiographic alveolar bone loss in subgroups of 14-year-old schoolchildren in Oslo. *Journal of clinical periodontology*, 1988; 15(2):130±3. PMID: 3162245
5. Ghijssels E, Coucke W, Verdonck A, Teughels W, Quirynen M, Pauwels M, et al. Long-term changes in microbiology and clinical periodontal variables after completion of fixed orthodontic appliances. *Orthodontics & craniofacial research*, 2014; 17(1):49±59.
6. Janson G, Bombonatti R, Brandao AG, Henriques JF, de Freitas MR. Comparative radiographic evaluation of the alveolar bone crest after orthodontic treatment. *American journal of orthodontics and dentofacial orthopedics: official publication of the American Association of Orthodontists, its constituent societies, and the American Board of Orthodontics*, 2003; 124(2):157±64.
7. Dahlen G. Microbiological diagnostics in oral diseases. *Acta Odontol Scand.* 2006;64:164–168.
8. Petti S, Barbato E, Simonetti D'Arca A. Effect of orthodontic therapy with fixed and removable appliances on oral microbiota: a six-month longitudinal study. *New Microbiol* 1997;20:55-62.
9. Faveri M, Figueiredo LC, Duarte PM, Mestnik MJ, Mayer MP, Feres M. Microbiological profile of untreated subjects with localized aggressive periodontitis. *Journal of clinical periodontology*, 2009; 36 (9):739±49. doi: 10.1111/j.1600-051X.2009.01449.x PMID: 19637996
10. Feng X, Zhang L, Xu L, Meng H, Lu R, Chen Z, et al. Detection of eight periodontal microorganisms and distribution of Porphyromonas gingivalis fimA genotypes in Chinese patients with aggressive periodontitis. *Journal of periodontology*, 2014; 85(1):150±9. doi: 10.1902/jop.2013.120677 PMID: 23646850
11. Elamin A, Albandar JM, Poulsen K, Ali RW, Bakken V. Prevalence of Aggregatibacter actinomycetemcomitans in Sudanese patients with aggressive periodontitis: a case-control study. *Journal of periodontal research*, 2011; 46(3):285±91. doi: 10.1111/j.1600-0765.2010.01337.x PMID: 21332472
12. Griffen AL, Becker MR, Lyons SR, Moeschberger ML, Leys EJ. Prevalence of Porphyromonas gingivalis and periodontal health status. *J Clin Microbiol.* 1998;36:3239–3242.
13. Socransky SS, Haffajee AD, Cugini MA, Smith C, Kent RL Jr. Microbial complexes in subgingival plaque. *J Clin Periodontol.* 1998;25:134–144.
14. Tanner AC, Maiden MF, Zambon JJ, Thoren GS, Kent RL Jr. Rapid chair-side DNA probe assay of Bacteroides forsythus and Porphyromonas gingivalis. *J Periodontol Res.* 1998;33:105–117.
15. Morinushi T, Lopatin DE, Van Poperin N, Ueda Y. The relationship between gingivitis and colonization by Porphyromonas gingivalis and Actinobacillus actinomycetemcomitans in children. *J Periodontol.* 2000;71:403–409.
16. Dahlen G. Microbiological diagnostics in oral diseases. *Acta Odontol Scand.* 2006;64:164–168.
17. Paolantonio M, Pedrazzoli V, di Murro C, di Placido G, Picciani C, Catamo G, De Luca M, Piaccolomini R. Clinical significance of Actinobacillus actinomycetemcomitans in young individuals during orthodontic treatment. A 3-year longitudinal study. *J Clin Periodontol.* 1997;24:610–617.
18. Leung NM, Chen R, Rudney JD. Oral bacteria in plaque and invading buccal cells of young orthodontic patients. *Am J Orthod Dentofacial Orthop.* 2006;130:698.e11–18.
19. Ristic M, Vlahovic Svabic M, Sasic M, Zelic O. Clinical and microbiological effects of fixed orthodontic appliances on periodontal tissues in adolescents. *Orthod Craniofac Res.* 2007;10:187–195.
20. Socransky SS, Haffajee AD, Cugini MA, Smith C, Kent RL Jr. Microbial complexes in subgingival plaque. *Journal of clinical periodontology*, 1998; 25(2):134±44. PMID: 9495612
21. Teixeira SR, Mattarazo F, Feres M, Figueiredo LC, de Faveri M, Simionato MR, et al. Quantification of Porphyromonas gingivalis and fimA genotypes in smoker chronic periodontitis. *Journal of clinical periodontology*, 2009; 36(6):482±7. doi: 10.1111/j.1600-051X.2009.01411.x PMID: 19508247
22. Braga RR, Carvalho MA, Bruna-Romero O, Teixeira RE, Costa JE, Mendes EN, et al. Quantification of five putative periodontal pathogens in female patients with and without chronic periodontitis by real-time polymerase chain reaction. *Anaerobe*, 2010; 16(3):234±9. doi: 10.1016/j.anaerobe.2010.02.007 PMID: 20193770

23. **Tomita S, Komiya-Ito A, Imamura K, Kita D, Ota K, Takayama S, et al.** Prevalence of Aggregatibacter actinomycetemcomitans, Porphyromonas gingivalis and Tannerella forsythia in Japanese patients with generalized chronic and aggressive periodontitis. *Microbial pathogenesis*, 2013; 61±62:11±5. doi: 10.1016/j.micpath.2013.04.006 PMID: 23608307
24. **Freitas AO, Marquezan M, Nojima Mda C, Alviano DS, Maia LC.** The influence of orthodontic fixed appliances on the oral microbiota: a systematic review. *Dental Press J Orthod*, 2014; 19(2):46±55. doi: 10.1590/2176-9451.19.2.046-055.oar PMID: 24945514
25. **Thornberg MJ, Riolo CS, Bayirli B, Riolo ML, Van Tubergen EA, Kulbersh R.** Periodontal pathogen levels in adolescents before, during, and after fixed orthodontic appliance therapy. *American journal of orthodontics and dentofacial orthopedics: official publication of the American Association of Orthodontists, its constituent societies, and the American Board of Orthodontics*, 2009; 135(1):95±8.
26. **Socransky SS, Haffajee AD.** The bacterial etiology of destructive periodontal disease. *J Periodontol* 1992;63:322-331.
27. **Friedman M, Harari D, Rax H.** Plaque inhibition by sustained release of chlorhexidine from removable appliances. *J Dent Res* 1985;64:1319-1321.
28. **Pender N.** Aspects of oral health in orthodontic patients. *Br J Orthod* 1986;13:95-103.
29. **Petti S, Barbato E, Simonetti D'Arca A.** Effect of orthodontic therapy with fixed and removable appliances on oral microbiota: a six-month longitudinal study. *New Microbiol* 1997;20:55-62.
30. **Taylor MG, McGorray SP, Durrett S, Pavlow S, Downey N, Lenk M, et al.** Effect of Invisalign aligners on periodontal tissues. *J Dent Res* 2003;82:1483.
31. **Jervøe-Storm PM, Koltzsch M, Falk W, Dörfler A, Jepsen S.** Comparison of culture and real-time PCR for detection and quantification of five putative periodontopathogenic bacteria in subgingival plaque samples. *J Clin Periodontol* 2005;32:778-783.
32. **Lau L, Sanz M, Herrera D, Morillo JM, Martin C, Silva C.** Quantitative real-time polymerase chain reaction versus culture: A comparison between two methods for the detection and quantification of Actinobacillus actinomycetemcomitans, Porphyromonas gingivalis and Tannerella forsythensis in subgingival plaque samples. *J Clin Periodontol* 2004;31:1061-1069.
33. **Jordan C, LeBlanc DJ.** Influences of VFB orthodontic appliances on oral populations of mutans Streptococci. *Oral Microbiol Immunol* 2002;17:65-71.
34. **Rudney JD, Chen R, Sedgewick GJ.** Intracellular Actinobacillus actinomycetemcomitans and Porphyromonas gingivalis in buccal epithelial cells collected from human subjects. *Infect Immun* 2001; 69: 2700–2707.
35. **Baehni PC, Guggenheim B.** Potential of diagnostic microbiology for treatment and prognosis of dental caries and periodontal diseases. *Crit Rev Oral Biol Med* 1996; 7: 259–277.
36. **Mayanagi G, Sato T, Shimauchi H, Takahashi N.** Detection frequency of periodontitis-associated bacteria by polymerase chain reaction in subgingival and supragingival plaque of periodontitis and healthy subjects. *Oral Microbiol Immunol* 2004; 19: 379–385.
37. **Zadeh HH, Nichols FC, Miyasaki KT.** The role of the cell-mediated immune response to Actinobacillus actinomycetemcomitans and Porphyromonas gingivalis in periodontitis. *Periodontol* 2000 1999; 20: 239–88.
38. **Okada M, Hayashi F, Nagasaka N.** Detection of Actinobacillus actinomycetemcomitans and Porphyromonas gingivalis in dental plaque samples from children 2 to 12 years of age. *J Clin Periodontol* 2000; 27: 763–768.
39. **Petti S, Barbato E, Simonetti D A** 1997 Effect of orthodontic therapy with fixed and removable appliances on oral microbiota: a six-month longitudinal study. *New Microbiology* 20: 55–62
40. **Choi D-S, Cha B-K, Jost-Brinkmann P-G, Lee S-Y, Chang B-S, Jang I, et al.** Microbiologic Changes in Subgingival Plaque After Removal of Fixed Orthodontic Appliances. *The Angle Orthodontist*, 2009; 79(6):1149±55. doi: 10.2319/111808-593R.1 PMID: 19852608
41. **Sandic MZ, Popovic B, Carkic J, Nikolic N, Glisic B.** Changes in subgingival microflora after placement and removal of fixed orthodontic appliances. *Srpski arhiv za celokupno lekarstvo*, 2014; 142(5±6):301± 5. PMID: 25033585
42. **Yanez-Vico RM, Iglesias-Linares A, Ballesta-Mударra S, Ortiz-Ariza E, Solano-Reina E, Perea EJ.** Short-term effect of removal of fixed orthodontic appliances on gingival health and subgingival microbiota: a prospective cohort study. *Acta odontologica Scandinavica*, 2015; 73(7):496±502. doi: 10.3109/00016357.2014.993701 PMID: 25631494
43. **Sallum EJ, Nouer DF, Klein MI, Goncalves RB, Machion L, Wilson Sallum A, et al.** Clinical and microbiologic changes after removal of orthodontic appliances. *American journal of orthodontics and dentofacial orthopedics: official publication of the American Association of Orthodontists, its constituent societies, and the American Board of Orthodontics*, 2004; 126(3):363±6.
44. **Thornberg MJ, Riolo CS, Bayirli B, Riolo ML, Van Tubergen EA, Kulbersh R.** Periodontal pathogen levels in adolescents before, during, and after fixed orthodontic appliance therapy. *American journal of orthodontics and dentofacial orthopedics: official publication of the American Association of Orthodontists, its constituent societies, and the American Board of Orthodontics*, 2009; 135(1):95±8.
45. **Kim K, Jung WS, Cho S, Ahn SJ.** Changes in salivary periodontal pathogens after orthodontic treatment: An in vivo prospective study. *Angle Orthod*, 2015.
46. **Krishnan V, Ambili R, Davidovitch Z, Murphy Neal C.** Gingiva and Orthodontic Treatment. *Seminars in Orthodontics*, 2007; 13(4): 257±71.
47. **Lee SM, Yoo SY, Kim H-S, et al.** Prevalence of putative periodontopathogens in subgingival dental plaques from gingivitis lesions in Korean orthodontic patients. *J Microbiol* 2005; 43: 260–265.
48. **Armitage GC.** Comparison of the microbiological features of chronic and aggressive periodontitis. *Periodontology* 2000, 2010; 53:70±88. doi: 10.1111/j.1600-0757.2010.00357.x PMID: 20403106
49. **Pan S, Liu Y, Zhang L, Li S, Zhang Y, Liu J, Wang C, Xiao S.** Profiling of subgingival plaque biofilm microbiota in adolescents after completion of orthodontic therapy. *PloS one*. 2017 Feb 3;12(2):e0171550.

50. **Cernochova P, Augustin P, Fassmann A, Izakovičová-Hollá L.** Occurrence of periodontal pathogens in patients treated with fixed orthodontic appliances. *Scr Med (Brno)*. 2008 Jun;81:85-96.
51. **Tamura K. and Nei M.** (1993). Estimation of the number of nucleotide substitutions in the control region of mitochondrial DNA in humans and chimpanzees. *Molecular Biology and Evolution* 10:512-526.
52. **Kumar S., Stecher G., and Tamura K.** (2016). MEGA7: Molecular Evolutionary Genetics Analysis version 7.0 for bigger datasets. *Molecular Biology and Evolution* 33:1870-1874.
53. **Kavarthapu Avinash and Kaarthikeyan Gurumoorthy,** 2016. "Knowledge of an orthodontist towards periodontal health care during orthodontic treatment", *International Journal of Current Research*, 8, (11), 41287-41290

