# Genotoxic Effects of Synthetic Fertilizer Urea in Channa punctatus

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*Abstract*: Urea, a common fertilizer used to supplement soil with nitrogen, induced 1.33%, 1.60%, and 1.83% of micronuclei and 21.6%, 24.0% and 26.0% of chromosomal abnormalities upon three different doses. The frequency of abnormalities increased with increase of doses. Polyploidy and aneuploidy were common among gross, while acentric fragments, minute fragments, chromatid breaks were more common among individual type of abnormalities. The individual type of damages were more prominent than gross type. This might be due to the formation of electrophilic radicals / ions during the metabolization of mutagens that attack the nucleophilic site of DNA leading to structural changes in chromosomes.

### Keywords: Urea, Genotoxicity, Micronuclei, Chromosomal abnormalities, Channa punctatus.

### 1. Introduction

The vast majority of teleost fishes are ammoniotelic excreting ammonia as the major nitrogenous end product in response to their aquatic habitat (Wood 1993; Saha and Ratha 1998). The ammonia stress on fishes includes accumulation of amino acid in different body tissues, (Levi et.al.1974; Das 1981; Dabrowska and Wlasow 1986 ;Iwata 1998;Saha 1992). Several organs such as gills, skin, liver, kidney, intestine, and gonads are extensively studied in fishes (Chatterjee and Bhattacharya 1983;Thurston et.al.1984; Ram and Sathyanesan 1986;1987a; Bhattacharya et.al 1989;Wright et.al. 1989; Banerjee and Paul 1993). Contamination of water bodies with ammonia takes place during application of the inorganic fertilizer, urea, ammonium sulphate(used for agriculture, aquaculture,)(Jhingran 1983;Ram and Sathyanesan 1987b;Sarkar 1991;Varadachari 1992) as well as ammoniotelic properties of fishes that aid also some amount of the nitrogen content in the aquatic medium.

Urea is a major nitrogen fertilizer to enrich the soil with nitrogen.Ammonia that emanates from the urea applied to agricultural fields, contribute to acid rain, while nitrates produced in soil contribute to contamination of ground water due to leaching of nitrates (http:/<u>www.fao.org/docrep/W2598E/w2598e 0.4</u> htm 6/22/2005).This fertilizer has been found to be present either in residual or some metabolised /derived form among the plants grown over them, and thus get accessed to the body of the animal that feed upon these plants (Baker and Chesnin 1975;Chaurasia and Sinha 1989;Current Science,Nov 2000).As agricultural run-off they pose a serious effect that induce various histopathological (Srivastava and Sravastava 1979;Nanda et.al, 2004;Ravindar kumar,2000) and cytogenetical changes (Jha,1998;Kohlpoth,1999;Dashwood, 1998;Baksi,1990)in the plants,aquatic animals,cattles and humans(Dravyam and Rajamanickam,2003,Neff,1985;Bhaskaran,1988;Singh et.al, 1998; Gupta, 2000, http://www 3. interscience wiley,.com/cgi-bin/abstract/ABSTRACT? CRETRY=1 &SRETRY=6/15/06).

Paddy fields are inhabited by some of the air-breathing fishes where pesticides and fertilizers are used regularly in greater amount. 40-45mg N/1 nitrate concentration has been reported in irrigated wells around the paddy fields.(http://www.fao.org/docrep/W2598E/w2598e 0.4 htm,6/22/2005).As nitrogen is a major component of chromosomes (in protein and DNA) it is quite possible that residues or metabolites of urea may cause some damages in fishes. The genotoxic effects of agrochemicals has been reported in various test system (Chaurasia and Sinha, 1987, 1990; Chaurasia 1991), but very few reports are available in the air-breathing fishes. So, the present investigation was therefore taken up to study the hitherto almost unknown genotoxic effect of urea on chromosomal abnormalities and incidence of micronuclei in Channa punctatus.

### 2. Materials and Method

Two test system viz; micronucleus test (from peripheral blood cells) and mitotic chromosomes from head kidney) were used.10-15 days acclimatized fishes were treated with freshly prepared doses of urea with three different concentration i.e Sub –lethal (SL-2.0%),half of the sub-lethal(HSL-1.0%) and quarter of the Sub-lethal (QSL-0.5%) for 7consecutive days. The animals were sacrificed after seven days of the termination of treatment.

The micronucleus test was conducted in peripheral blood cells. A film of blood smear was prepared after mixing with few drop of anticoagulant (0.1% trisodium citrate solution) on a greese free clean slide. Preserved in methanol for 10 minutes , stained with 0.15% Leishman's stain for 20-25 minutes and cleared in xylene for 5 minutes .3000 RBC cells were screend. A concurrent control were carried out were animals were kept in fresh water.

For studying the chromosomal abnormalities,tissue from head kidney were taken and the slides were made by the conventional Colchicine –hypotonic –acetoalcohol-flame drying –giemsa staining techinuqe.300 well spread and randomly selected metaphase plates were screened and data were analyzed by statistical procedure. A seperate common control was also carried out. 3. Results and discussion

Amidst 3000 RBCs,only 0.43% micronuclei were found in the control group while 1.33%, 1.60% and 1.83% micronuclei upon three doses of urea (SL,HSL& QSL)were observed (Table -1).A close observation of data revealed that the effect was dose –dependent (Graph -1).Most of the cells were found to have only one micronucleus of very small size (due to acentric fragment )or bigger size (due to lagging of whole chromosome )but very few cells were found to have more than one micronucleus.

Amidst 300 metaphase plates,21.6%,24.0% and 26.0% chromosomal abnormalities were found upon treatment with three doses of urea in contrast to 5.33% in the control (Table -2).The abnormalities that were found can be put in two categories – gross and individual ones.The insignificant gross changes were the stickiness, polyploidy , hypoploidy etc..The significant individual changes were mostly breaks in the chromosomes (Chromatid break, chromatid gap).Acentric fragment and minute fragment were also observed that might be due to breaks and deletion of certain part of chromosomes (telomeric or interstitial part ).A quantitative estimation revealed that the abnormalities increased with the increase of the doses. Thus the effect was dose dependent (Graph-1).The individual type of damages were more prominent than the gross type because urea is synthetic in nature. While Chaurasia and Sinha 1987,1988, 1990),Chaurasia et.al. (2005)were studying on genotoxicity induced by fertilizer and silk dyeing wastes; Kumar and Sinha(1989)on doses –dependent genotoxic effects of synthetic pesticides,they observed that the individual type of damages induced more gross type of abnormalities than individual types.This differential sensitivity might be occurred at two different levels. First ,the damages at protein level either on spindle protein or on protein packing. Second , by the production of electrophilic ions and reactive radical during the metabolization of mutagens (Klopman et.al..1985).Such electrophilic reactive radicals /ions might attack to nucleophilic site of DNA leading to structural changes in chromosomes(Awasthy et.al.1999).

The result thus shows that the synthetic fertilizer urea was mutagenic and harmful to the fishes with a regular deterioration of their population and thus affecting the economy of our country.

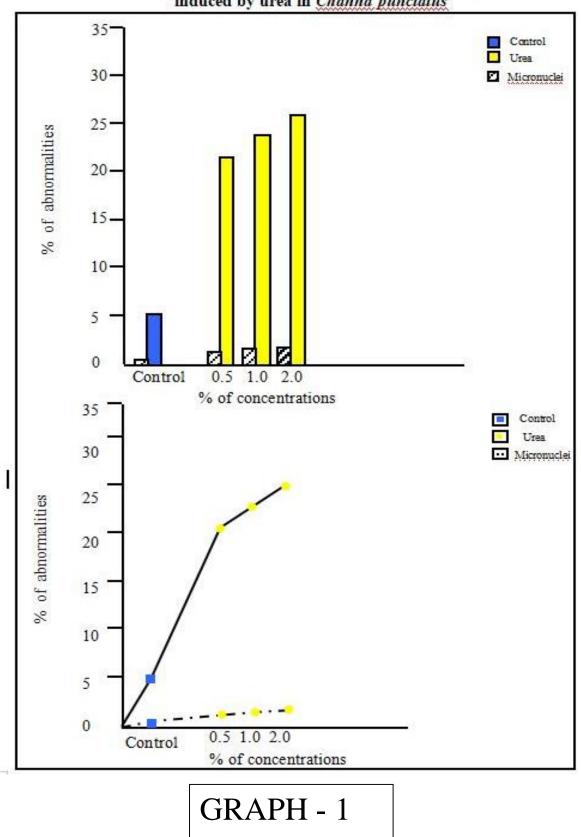
Incidence of micronuclei(N=3000) after urea treatment in channa punctatus.

EXPERIMENT	Abnormal cells Micronuclei
TREATMENT (in %	
Control 0.5	13 $0.43 \pm 0.11$ 13 $0.43 \pm 0.11$ 40 $1.22 \pm 0.20*$ 40 $1.22 \pm 0.20*$
0.5	$\begin{array}{rrrrrrrrrrrrrrrrrrrrrrrrrrrrrrrrrrrr$
2.0	$\begin{array}{cccccccccccccccccccccccccccccccccccc$
2.0	Table-1

Table-2	$2.0\%   300   77   25.6 \pm  2.51*   7   2.33 \pm  0.87   73   24.3 \pm  2.47*   80   26.0 \pm  2.53*$	1.0%	$0.5\% \qquad 300 \qquad 64 \qquad 21.3 \qquad \pm \qquad 2.36^*  7  2.33  \pm  0.87  58  19.3  \pm  2.27^* \qquad 65  21.6  \pm  2.37^*$	Control 300 15 5.00 $\pm$ 1.25 1 0.33 $\pm$ 0.33 15 5.00 $\pm$ 1.25 16 5.33 $\pm$ 1.29	$\begin{tabular}{lllllllllllllllllllllllllllllllllll$	C		$\begin{array}{r cccccccccccccccccccccccccccccccccccc$	Gross Abnormality No. % ± S.E 1 0.33 ± 0.33 8 2.66 ± 0.92 7 2.33 ± 0.87 7 2.33 ± 0.87 7 2.33 ± 0.87	S.E 1.25 2.36* 2.46* 2.51*	Abnorn metaph 5.00 21.3 24.0 25.6		nt No. o. metaphas analyzec 300 300 300 300	eatmer 5% 0%
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Chromosomal abnormalities(N=300) after urea treatment in channa punctatus

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Comparative graph of micronucleus and chromosomal abnormalities induced by urea in *Channa punctatus* 

#### References

Awasthy K.S, Chaurasia O.P and Sinha S.P. 1999 : Prolonged Murine Genotoxic effects of crude Extracted from Neem. *Pytotherapy Research*. **13**:81-83.

Awasthy K.S, Chaurasia O.P, Sinha S.P. and Khan P.K 2000.Differential genotoxicity of the crude leaf extract of medicinal plant, Casearia tomentosa, *Biomedical and environmental Science*. **13** : 12 - 18.

Baker D.E. and Chesnin, L. (1975). Chemical monitoring of soils for environmental quality and human health. *Adv. in Agronomy* **27**:306-366.

Baksi S.M, Frazier J.M, (1990) isolated fish hepatocytes model systems for toxicology research, Aquat. Toxicol. 16, 229-256.

Banerjee TK and Paul VI 1993.Estimation of acute toxicity of ammonia sulphate to the fresh water cat fish Heteropneustes fossilis 11. A histopathological analysis of the epidermis : *Biomed.Environ*. Sci. **6** 45-48

Bhaskaran R,1988 Effect of DDT and methyl parathion on the mitochondrial respiration SDH and ATpase activity of an airbreathing fish, *Channa straitus.Environ and Ecol.***6** : 198.

Bhattacharya T, Bhattacharya S, Ray A K and Dey S 1989 influence of industrial pollutants on thyroid function in *Channa punctatus* (bloch); *Indian.J.Exp.Biol.*27 65-66.

Bose S and Sinha S.P. 1994 : Modulation of Ochratoxin produced enotoxicity by Vitamin C in mice. *Fd. Chem Toxicol.* .32 : 533 - 537

Chaurasia O.P and Sinha S.P.1989 Cytogenetic effects of urea on onion root tip cells:Perspectives in *Cytology and genetics* **6**: 715-718

Chaurasia O.P, and Sinha, S.P 1987 Effect of urea on mitotic chromosome. of mice and onion. Cytologia 52: 877-882.

Chaurasia O.P and. Sinha S.P 1988 Induction of mitotic chromosome anomalies in mice by single super phosphate. *Cytologia*. **53** 485-489.

Chaurasia O.P and Sinha S.P 1990 Induction of mitotic chromosome anomalies in mice by muriate of potash. *Cytologia* .55 87 - 90.

Chaurasia O.P (1991) Randomness of chromosomes breaks in bone marrow cells of fertilizer fed mice, *Mus musculus.Cytobios* . 67 (268), 7-12.

Chaurasia O.P, Kumar A, Kumari M (2005) Genotoxic effect of silk dyeing wastes in Bone marrow cells of mice, *Mus musculus*. *Cytologia*.**70**(4): 381-385

Chatterjee S and Bhattacharya S 1983. Ammonia induced changes in the hepatic glutathione level of an air-breathing fresh water teleost *Channa punctatus* (Bloch); *Toxicol .Lett.* **17** 329 – 333.

Dabrowska H and Walsow T 1986 .Sub lethal effect of ammonia and certain biochemical and haematological indicators in common crap(cyprinus carpio L);*Comp.Biochem physiol*.C83 179-184.

Das N P 1981.changes in nitrogen metabolism of *Channa punctatus* (Bloch) due to ambient ammmonia,Ph.d thesis,Berhampur University Berhampur

Dashwood R.H, Bailey G.S, 1998 Use of fish and fish transgenics in laboratory and field genotoxicological studies, *Mutat.Res.***399** 123-124.

Dharmshila K. and Sinha S.P.1994.Effect of retinol on ochratoxin produced genotoxicity in mice.*Fd. Chem Toxicol..***32**: 471-475.

Gupta V.K, Sharma JP, Verma AK 2000, (Rural Dev Unit, Regl Res Lab(CSIR), Canal Rd, Jammu (180001). Effect of some fertilizers on the early developmental stages of *Cyprinus carpio Linn. Poll Res*,**19**(3)369-375.

http: // www 3. interscience wiley. com / cgi - bin / abstract?

CRETRY =1 & SRETRY=6/15/2006.Micronucleus test in fish cells.A bioassay for in situ monitoring of Genotoxic pollution in the marine environment.

Introduction to agricultural water pollution Page 1 of 19(http://www.fao.org / docrep / W2598E / w2598e 0.4 htm)-6 /22 /2005.

Iwata K 1988.Nitrogen metabolism in the mudskipper, Periopthalmus cantonens : Changes in free amino acids & related compounds in various tissues under condition of ammonia loading with special references to high ammonia tolerance ; *Comp. Biochem Physiol* ; A**91** 499 - 508.

Jha A.N, 1998 use of aquatic invertebrates in genotoxicological studies, Mutat.Res. 399 1-2

Jhingran V G 1983 fish and fisheries of India (New Delhi : Hindustan Publishing Corporation) PP 1-666.

Klopman G, Conttreras R, Reosenkranz H.S and Waters M.D, 1985 Structure-genotoxic activities relationship of pesticides: Comparisons of the results from several short term assays *Mutat.Res*.147:343-356.

Kohlpoth M, Rusche B, Nusse M, (1999) Flow cytometric measurement of micronuclei induced in a permanent fish cell line as a possible screening test for the genotoxicity of industrial waste waters, *Mutagenesis* 14, 397-402.

Kumar D and Sinha S.P. 1989. Threshold dose of cytogenetic toxicity of linadane, malathion and metacid in Alluim cepa root tip cells *Cytologia* **54**: 547-552.

Kumar R (PG Dept Zoo, SSV (PG) Coll, Hapur-245101) 2000, .Chronic ammonia induced histopathological changes in Indian subtropical fresh water murrel *channa punctatus* (Bloch) *Polln Res*, **19** (4) 611 - 613.

Levi G, Morisi G, colleti A and Catanzaro R 1974.Free amino acid in fish brain : normal levels and hanges upon exposure to high ammonia concentration in viva and upon in cubation of brain slices Comp *Biochem. Physiol*.A49 623-636.

Nanda P, Panigrahi S, Nanada BR, Behera M.K and Kriali E, (2004) Histopathological abnormalities in the fish climbing perch *Anabas testudineus* due to paper mill effluents. *Env. Eco.***22** 24-25.

Neff J.M, 1985 Use of biochemical measurement to detect pollutant mediated damage to fish *ASTM.Spec.Tech,Publ.*,**854**:155-183 Prakash Rao.E.V.S and Puttana K.2000.Nitrates,agriculture and environment: *Current Science*.Vol.**79** No.9 10 November 2000.

Ram R N and Sathyanesan A G 1986 Ammonium sulphate induced nuclear changes in the oocyte of the fish, Channa punctatus (Bl.); *Bull. Environ. Contam.Toxicol.***36** 871-875.

Ram R N and Sathyanesan A G 1987a.Effect of Chronic Exposure of commercial nitrogenous fertilizer, ammonium sulphate on testicular development of a teleost Channa punctatus (Bloch)*Indian J*.*Exp.Biol* **25** 667-670.;

Ram R Nand Sathyanesan A G 1987b.Histopathological changes in liver and thyroid of the teleost fish, Channa punctatus (Bloch) in response to ammonium sulphate fertilizer treatment; ecotoxical.*Environ.Safety*. **13** 185-190

241

Saha N and Ratha B K 1998.Ureogenesis in Indian air-breathing teleosts:adaption to environmental constrains, *comp. biocom. physiol.* A120 195-208.

Saha T K 1992 Effect of a Sub-lethal concentration of ambient ammonia-nitrogen on protein metabolism of *Channa punctatus* (Bloch), Ph.D. thesis, Visva-Bharti University ,Santiniketan.

Sarkar S k 1991.Use of ammonium sulphate nitrate in rearing major carp Spawn; Geobios 18 177-181.

Selvarani D and Rajamanickam C, 2003 Toxicity of PCB 1232 on mitochondria of fish *Arius Caelatus* (Valenciennes).*I. J. Expt. Biol.* **41**, 336-340.

Singh T.P, Lal Bechan and Yadav A.K, 1998 Pesticides and Fish In: Pesticides, Man and Biosphere(Ed) Shukla Pp 265-319

Srivastava, G.J and Srivastava, O.P(1979) Urea induced histopathology in skin and gastric lining of the teleost, *Channa punctatus* (B1) *Indian J.Exp.Biol.***17**, 840-843.

Thurston RV, Russo RC, Leudtke RJ, Smith CE, Meyn EL, Chakoumakos C, Wang KC and Brown CJD 1984. Chronic toxicity of ammonia on rainbow trout; trans. an. *Fish. Soc.* **113** 56 -73.

Varadachari C 1992. Phosphoric acid, phosphates and fertilizers for the future, Proc. Indian, Natl. Sci. Acad. B 58 119-126.

Wood CM 1993 Ammonia and Urea metabolism and cxcretion in the phyciology of fishes (ed.) DH Evans (Boca Raton :CRC Press) PP 379-425.

Wright P A, Randall D J and Perry II SF 1989 Fish gill water boundary layer: A site of linkage between carbon dioxide and ammonia excretion, *J.Comp.Physiol.* **158** 627-635.

