

A Review on the green synthesis, and application of silver nanoparticles

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Abstract - Nanoparticles deals with particles having size in the range 1nm-100nm. Nanoparticles of noble metals such as silver show distinct physical, chemical and biological properties which vary according to their varying sizes and are also significantly different from the bulk silver materials. Their unique properties are due to their extremely small particle dimension, high surface area, size quantization effect and several other factors. Silver nanoparticles exhibit electromagnetic, optical, catalytic properties and these properties are highly influenced by the shape and size distribution of the nanoparticles. Silver nanoparticles of different size and shapes can be synthesized by tuning the various synthetic routes. Silver nanoparticles are recognized for its widespread applications also. So this review deals with the green synthetic procedures and applications of silver nanoparticles in medicine.

Keywords- Silver nanoparticles, Green reduction method, Antibacterial activity.

1. INTRODUCTION

Nanotechnology is an emerging field of science which involves the synthesis and development of various nanomaterials [1]. It can be defined as a whole knowledge on fundamental properties of nano-sized objects[2-4].The term 'nano' indicates one billionth or 10^{-9} units. It is universally accepted that nanoparticles are clusters of atoms in the size range of 1-100nm.[5]

Nanoscience has been established recently as a new interdisciplinary science.[6] At present time nanochemistry becomes one of the main growing directions of nanoscience[4]. Advances in this field largely depend on the ability to synthesize nanoparticles of various nano materials, based on their sizes, and shapes, as well as their efficiency to assemble them into complex architectures [7]. Nanotechnology provides the ability to engineer the properties of materials by controlling their size, and this has driven research towards a multitude of potential uses for Nanomaterial [8].

With the evolution of nanomedicine as a study for treating infections, metallic silver in the form of nanoparticles has regained its significance [9,10]. Several bacteria have developed resistance against antibiotics, which has challenged the treatment of human infections[11-13]. Therefore, silver nanoparticles (AgNPs) as an antimicrobial agent seems to be beneficial compared to antibiotics[14].

Silver nanoparticles have been the prime focus of the nanoparticles' research industry due to their unique thermal [15], electrical [16] and optical [17] properties and also because of the use of these structures in products that range from photovoltaics [18] to biological and chemical sensors [19]. In recent years nanoparticles of silver have been found to exhibit interesting antibacterial activities [20,21].Antibacterial activity of the silver-containing materials can be used, for example, in medicine to reduce infections as well as to prevent bacteria colonization on prostheses[22], catheters[23,24],vascular grafts[25], dental materials[26], stainless steel materials[27] and human skin[26,28]. Contrary to bactericide effects of ionic silver, the antimicrobial activity of colloid silver particles are influenced by the dimensions of the particles, the smaller the particles, the greater the antimicrobial effect [20].The catalytic activity of the silver nanoparticles is greatly ruled by their size, structure,shape,size distribution and chemical- physical environment. Thus proper control of the shape and size distribution of these nanoparticles is necessary. The specific control of shape, size, and size distribution is achieved by varying the synthetic strategies, reducing agents and stabilizers[6].

Biosynthesis of silver nanoparticles using plant extracts may be influenced directly or indirectly by photochemical in extracts such as phenols, flavonoids and antioxidants as well as the physicochemical factors governing the kinetics of the reactions.This route is preferably docile as it is ecofriendly, involves less energy intensive processes and is cost effective. Moreover, it is an efficient way of waste biomass utilization for the biosynthesis of silver nanoparticles. Currently silver nanoparticles are prepared by different methods including electrolysis, physical, chemical and biological methods [29, 30]. The bio reduction of silver ions to yield silver nanoparticles seems to be an efficient, cost effective and eco friendly approach. Biosynthesis of silver nanoparticles using the plant extracts of *Lippia citriodora* [31], *Citrus sinensis* [32], *Magnolia kobus* [33], neem leaf, geranium leaf [34] have already been reported.

2. GREEN SYNTHESIS OF SILVER NANOPARTICLES

2.1 Green synthesis of silver nanoparticles using plant parts

The use of plant parts for the synthesis of silver nanoparticles has drawn attention because of its rapid, eco-friendly, cost-effective, easy approach for biosynthetic processes. The reduction and stabilization of silver ions by combination of biomolecules such as proteins, amino acids, enzymes, polysaccharides, alkaloids, tannins, phenolics, saponins, terpenoids and vitamins which are already established in the plant extracts having medicinal values and are environmental benign, yet chemically complex structures[35].

In this procedure the various parts of the plant such as fresh leaves, stem, fruit, fruit peel, seed etc are finely cleansed and boiled in 100mL distilled water contained in conical flask. This plant extract was then added to 1mM silver nitrate solution and heated at about 80°C for few hours. Change in colour of the solution to reddish brown will be observed during the heating process after 15-20 mins. The resulting solution then needs to be centrifuged at 10,000 rpm for about 25mins. The centrifuged sample was then collected and vacuum dried for analysis[36-39]. These plant extracts contain biomolecules, which act as both reducing and capping agents that form stable and shape-controlled nanoparticles. Main compounds which affect the reduction and the capping of the nanoparticles are biomolecules such as phenolics, terpenoids, polysaccharides, flavones, alkaloids, proteins, enzymes, amino acids, and alcoholic compounds. However, quinol and chlorophyll pigments, linalool, methyl chavicol, eugenol, caffeine, theophylline, ascorbic acid, and other vitamins have also been reported. The nontoxic phytochemicals including aforementioned flavonoids and phenols have unique chemical power to reduce and also effectively wrap nanoparticles, thus preventing their agglomeration. Phenolic compounds possess hydroxyl and carboxyl groups, which are able to bind to metals.[40]

Table 1. Green synthesis of silver nanoparticles by different researchers using plant extracts.[41]

Plants	Size (nm)	Plant's part	Shape
<i>Alternanthera dentate</i>	50–100	Leaves	Spherical
<i>Acorus calamus</i>	31.83	Rhizome	Spherical
<i>Boerhaavia diffusa</i>	25	Whole plant	Spherical
<i>Tea extract</i>	20–90	Leaves	Spherical
<i>Tribulus terrestris</i>	16–28	Fruit	Spherical
<i>Cocous nucifera</i>	22	Inflorescence	Spherical
<i>Abutilon indicum</i>	7–17	Leaves	Spherical
<i>Pistacia atlantica</i>	10–50	Seeds	Spherical
<i>Ziziphora tenuior</i>	8–40	Leaves	Spherical
<i>Ficus carica</i>	13	Leaves	–
<i>Cymbopogon citratus</i>	32	Leaves	–
<i>Acalypha indica</i>	0.5	Leaves	–
<i>Premna herbacea</i>	10–30	Leaves	Spherical
<i>Calotropis procera</i>	19–45	Plant	Spherical
<i>Centella asiatica</i>	30–50	Leaves	Spherical
<i>Argyreia nervosa</i>	20–50	Seeds	–
<i>Psoralea corylifolia</i>	100–110	Seeds	–
<i>Brassica rapa</i>	16.4	Leaves	–
<i>Coccinia indica</i>	10–20	Leaves	–
<i>Vitex negundo</i>	5 & 10–30	Leaves	Spherical & fcc
<i>Melia dubia</i>	35	Leaves	Spherical
<i>Portulaca oleracea</i>	<60	Leaves	–
<i>Thevetia peruviana</i>	10–30	Latex	Spherical
<i>Pogostemon benghalensis</i>	>80	Leaves	–
<i>Trachyspermum ammi</i>	87, 99.8	Seeds	
<i>Swietenia mahogani</i>	50	Leaves	
<i>Musa paradisiacal</i>	20	Peel	
<i>Moringa oleifera</i>	57	Leaves	
<i>Garcinia mangostana</i>	35	Leaves	
<i>Eclipta prostrate</i>	35–60	Leaves	Triangles, pentagons, hexagons
<i>Nelumbo nucifera</i>	25–80	Leaves	Spherical, triangular

Plants	Size (nm)	Plant's part	Shape
<i>Acalypha indica</i>	20–30	Leaves	Spherical
<i>Allium sativum</i>	4–22	Leaves	Spherical
<i>Aloe vera</i>	50–350	Leaves	Spherical, triangular
<i>Citrus sinensis</i>	10–35	Peel	Spherical
<i>Eucalyptus hybrid</i>	50–150	Peel	
<i>Memecylon edule</i>	20–50	Leaves	Triangular, circular, hexagonal
<i>Nelumbo nucifera</i>	25–80	Leaves	Spherical, triangular
<i>Datura metel</i>	16–40	Leaves	Quasilinear superstructures
<i>Carica papaya</i>	25–50	Leaves	
<i>Vitis vinifera</i>	30–40	Fruit	

2.2 Green synthesis of silver nanoparticles using bacteria

Synthesis of Silver nanoparticles by microbes is due to their defense mechanism (resistance mechanism), and this is how the nanoparticles produced are useful to us. The resistance caused by the bacterial cell for silver ions in the environment is responsible for its nanoparticles synthesis. The silver ions in nature are highly toxic for the bacterial cells. So their cellular machinery helps in the conversion of reactive silver ions into stable silver atoms. The nanoparticles can be artificially synthesized in vitro using chemical method via ethanol. But, here the synthesis was done through *E. coli* under room temperature. The supernatant was taken from the nutrient broth, incubated overnight inoculated with *E. coli*. Then 1 mM of AgNO₃ (1% v/v) was added to the supernatant. The formation of silver nanoparticles was observed within 10 minutes. The color change was noticed from fine yellow color to reddish brown with time. [42]

2.3 Green synthesis of silver nanoparticles using fungi

Similar to bacteria, due to their tolerance and metal bioaccumulation ability, high binding capacity, and intracellular uptake, fungi have been of interest in biological production of the metallic nanoparticles. Compared to bacteria, fungi are simpler to handle in a laboratory process. The mechanism of nanoparticle production using fungi is different; fungi secrete large amounts of enzymes which are used to reduce silver ions that induce the formation of the metal nanoparticles [40].

3. BIOMEDICAL APPLICATIONS OF SILVER NANOPARTICLES

3.1 Antibacterial effects

Silver nanoparticles have antibacterial properties and can inhibit the reproduction of bacteria, which is a microbe. The silver nanoparticles can “inactivate proteins, blocking respiration and electron transfer, and subsequently inactivating the bacteria” [43]. Inactivation of the bacteria does not allow them to reproduce. The nanoparticles are able to interact with the microbes because the “cell wall peptidoglycans contain negatively charged molecules that will likely interact electrostatically with the silver ions” [44]. The silver nanoparticles naturally have a positive charge, which causes them to be attracted to the negatively charged molecules within the cell wall and causes damage to the cell by interrupting its natural processes. The antibacterial properties of the silver nanoparticles depend on the size of the particles; the smaller the size the greater the effect. The particle size is a major factor because the smaller the particle the greater the surface area, which allows for greater interaction with the bacteria. “Nanoparticles and silver ions interact with sulfur-containing compounds found in bacterial membrane protein and with phosphorous-containing compounds, such as DNA” [43]. The interaction with the DNA can also cause a decrease in microbe reproduction, allowing the antimicrobial effects on surfaces to be successful. [45]

3.2 Antiviral effects

Metal nanoparticles have been studied for their antimicrobial potential and have proven to be antibacterial agents against both Gram-negative and Gram-positive bacteria. Theoretically, any metal could be analysed for antiviral activity, however, little effort has been done to determine the interactions of metal nanoparticles with viruses, and only recently some studies have emerged showing that metal nanoparticles can be effective antiviral agents against HIV-1, hepatitis B virus, respiratory syncytial virus, herpes simplex virus type 1 monkeypox virus, influenza virus and Tacaribe virus [46]

The antiviral effects of Ag-NPs, most publications have suggested that Ag-NPs could bind to outer proteins of viral particles, resulting in inhibition of binding and the replication of viral particles in cultured cells. Although the antiviral mechanism of Ag-NPs has not been fully known yet, Ag-NPs are still suggested as potential antiviral agents in the future [47].

3.3 Anti-fungal effects

Kim *et al* studied the antifungal activities of Ag-NPs against a total of 44 strains of six fungal species from clinical isolates and ATCC strains of *Trichophyton mentagrophytes* (*T. mentagrophytes*) and *Candida albicans* (*C. albicans*). Results showed 80% inhibitory concentration (IC₈₀) from 1 to 7 µg ml⁻¹. The antifungal activity of Ag-NPs against *C. albicans* could be exerted by disrupting the structure of the cell membrane and inhibiting the normal budding process due to the destruction of the membrane integrity. Roe *et al* have tested the antifungal activity of plastic catheters coated with Ag-NPs (~100 nm thick), and results showed that the growth inhibition was almost complete for *C. albicans*. Pamacek *et al* investigated the antifungal activity of Ag-NPs prepared by the modified Tollens process. Results also revealed the minimum inhibition against *C. albicans* growth at 0.21 mg l⁻¹ using naked Ag-NPs, and 0.05 mg l⁻¹ using Ag-NPs modified with sodium dodecyl sulfate (SDS). Additionally, Ag-NPs effectively inhibited the growth of the tested yeasts at the concentrations below their cytotoxic limit against the tested human fibroblasts determined at a concentration equal to 30 mg l⁻¹ of Ag-NPs. Other publications also reported MICs of Ag-NPs from 0.4 to 3.3 µg ml⁻¹ against *C. albicans* and *C. glabrata* adhered cells and biofilm, and at 10 µg ml⁻¹ against *Trichophyton rubrum* (*T. rubrum*). In summary, Ag-NPs have also been revealed as potential biocide against fungal strains, and could help to prevent fungal infections for protection of human health[47].

3.4 Anticancer effects

Shahnaz Majeed et.al (2010) reported anticancer analysis (via MTT assay) efficacy of AgNPs on MCF-7 cancer cells. These AgNPs showed a concentration-dependent anticancer effect on MCF-7 cells. The IC₅₀ value was 70 µg/ml after 24 h of incubation. Upon further incubation for 48 h, the toxicity increased and the IC₅₀ value was 50 µg/ml. Thus, these nanoparticles display anticancer activity against MCF-7 breast cancer cells[47].

The cytotoxicity of the AgNPs were studied in vitro against HCT-116, MCF-7, Hep-G2 and Caco-2 cancer cell lines at different concentration (0, 5, 12.5, 25, 50 µg/mL).

The results obtained from MTT assay after 48hrs of incubation showed that fruits/AgNPs showed significant effect on Hep-G2 and MCF-7 with IC₅₀ = 17.2 and 22.4 µg respectively. Fruits/ AgNPs cytotoxic effect on HCT-116 and Caco-2 showed insignificant anticancer activity with IC₅₀ > 30µg/ml.

Leaves/SNPs was effective only against Hep-G2 cancer cell line, as it led to inhibition in cell growth as concluded from IC₅₀ values 10.2µg /mL

Treatment of tested cell cancer cell lines with seeds/AgNPs led to insignificant cytotoxic effect with IC₅₀ values above 30 µg.

At the same time treatment of HCT-116 and Hep-G2 cancer cell lines with roots/AgNPs led to significant cytotoxic effects with IC₅₀ 21.2 µg for HCT-116 and IC₅₀ = 22.4 µg for Hep-G2 . Insignificant cytotoxic effect was observed on MCF-7 and Caco-2 cell lines [48]. Thus silver nanoparticles exhibit significant anticancer effects against some human cancer cell lines.

CONCLUSION

The study of silver nanoparticles has emerged as a current trend due to its widespread applications. These nanoparticles have many important applications that include spectrally selective coating for solar energy absorption, as optical receptors, polarizing filters, catalysts in chemical reaction, and other biomedical applications. Thus green chemistry provides us an environment friendly, easy, cost effective and energy efficient single step technique for the synthesis of silver nanoparticles with no hazardous byproducts.

REFERENCES

- [1] Y.Li, P. Leung, L. Yao, Q.W. Song and E. Newton, Journal of Green chem,2007; 9: 852-858.
- [2] G. Sergeev, Nanochemistry 2006,Elsevier.
- [3] G.Sergeev, J. Nanoparticle Res.,Volume 5,2003,p.529
- [4] G.Sergeev, T.Shabatina, Colloids Surf A : Physicochem. Eng. Aspects,Volume 313, 2008, p.18
- [5] D.Williams, Biomaterials, Volume 29, 2008, p. 1737
- [6] M.M.Kholoud, Abou El-Nour, Ala'a Eftaiha, Abdulrhman Al-Warthan, Reda A.A.Ammar, Arabian Journal of Chemistry,Volume 3(3),2010,p. 135-140.
- [7] D.David, J.Evanoff and C.George, Chem. Phys. Chem,2005; 6: 1221 – 1231.
- [8] N.Saifuddin ,C.W. Wong and Nur Yasumira A.A., E-Journal of Chemistry,2009; 6(1): 61-70.Govindasamy Rajakumar & Sampath Marimuthu & Asokan Bagavan & Chidambaram Jayaseelan & Abdul Abduz Zahir & Gandhi Elango & Chinnaperumal Ka
- [9] Y. He, Z. Du, H. Lv et al., Int J Nanomedicine, 2013;8:1809–1815.
- [10]M. Rai, A. Yadav and A. Gade, Biotechnol Adv. ,2009; 27(1):76–83.
- [11]M.R. Shakibaie, P. Dhakephalkar, B.P. Kapadnis ,B.A. Chopade, Can J Microbiol.,1999;45(12):995–1000.
- [12]A.E.Waters, T. Contente-Cuomo , J. Buchhagen et al., Clin Infect Dis.,2011; 52(10):1227-1230.

- [13] P.K. Dhakephalkar, B.A. Chopade, *Biometals*, 1994; 7(1): 67-74.
- [14] I. Chopra, *J Antimicrob. Chemother.*, 2007; 59(4): 587-590.
- [15] K.S. Moon, H. Dong, R. Maric, S. Pothukuchi, A. Hunt, Y. Li and C.P. Wong, *J Electron Mater*, 2005; 34:168-175.
- [16] D. Chen, X. Qiao, X. Qiu and J. Chen, *J Mater Sci*, 2009; 44:1076-1081.
- [17] K.L. Kelly, E. Coronado, L.L. Zhao, G.C. Schatz, *J Phys Chem B*, 2003; 107:668-677.
- [18] W.J. Yoon, K.Y. Jung, J. Liu, T. Duraisamy, R. Revur, F.L. Teixeira, and P.R. Berger, *Sol Energy Mater Sol Cells*, 2010; 94:128-132.
- [19] A.D. McFarland A.D., R.P. Van Duyne R.P., *Nano Lett*, 2003; 3:1057-1062.
- [20] A.R. Shahverdi, A. Fakhimi, H.R. Shahverdi, M.S. Minaian, *Nanomedicine: Nanotechnology, Biology and Medicine*, 2007; 3(2): 168-171.
- [21] S. Pal S., Y. Kyung, J. Myong Song, *J Appl Environ Microbiol*, 2007; 73:1712-1720.
- [22] M.V. Roldán, A.L. Frattini, O.A. Sanctis, N.S. Pellegrini, *Anales AFA*, 2005; 17: 212-217.
- [23] H. Yin, T. Yamamoto, Y. Wada, S. Yanagida, *Materials Chemistry and Physics*, 2004; 83 :66-70.
- [24] Z. Zhu, L. Kai, and Y. Wang, *Materials Chemistry and Physics*, 2006; vol. 96, no. 2-3, pp. 447-453.
- [25] A.S. Edelstein A.S., R.C Cammarata R.C. (eds.). *Nanomaterials: synthesis, properties and applications*, Bristol and Philadelphia Publishers, Bristol, 1996.
- [26] M. Maillard, S. Giorgio, and M.P. Pileni, *Adv. Mater.*, 2002; 14(15): 1084-1086.
- [27] J.J. Mock, M. Barbic, D.R. Smith, D.A. Schultz and S. Schultz, *J. Chem. Phys.*, 2002; 116(15):6755- 6759
- [28] N. Duran, P.L. Marcato, O.L. Alves, and G.I. De Souza., *J. Nanobiotechnology*, 2005; 3: 1-7.
- [29] S.P. Dubey, M. Lahtinen, M. Sillanpaa, *Process Biochem*, 2010; 45: 1065-1071.
- [30] P. Usha Rani, R. Reddy, *Coll Surf A Physicochem Eng Asp*, 2011; 389: 188-194.
- [31] D. Cruz, P.L. Fale, A. Mourato, P.D. Vaz, M.L. Serralheiro and A.R. Lino, *Coll Surf B Biointerface*, 2010; 81: 67-73.
- [32] S. Kaviya, J. Santhanalakshmi, B. Viswanathan, J. Muthumary, K. Srinivasan, *Spectrochim Acta A Mol Biomol Spectrosc*, 2011; 79: 594-598.
- [33] B.S. Kim, S.H. Lee, B.K. Salunke, *Biotechnol Bioprocess Eng*, 2014; 19: 169-174.
- [34] P.C. Nagajyoti, T.N.V.K.V. Prasad, T.V.M Sreekanth and KapDuk Lee., *Digest J. Nanomat. Biostruct.*, 2011; 6(1):121-133.
- [35] N. Kulkarni, U. Muddapur, *Biosynthesis of Metal Nanoparticles: A Review. J. Nanotech.* 2014; 1-8.
- [36] S. Thirunavukkarasu, AR Abdul, R Govindasamy, M Sampath, B Asokan, J Chidambaram, AZ Abdul, E Gandhi, K Chinnaperumal, *Parasitol Res* 2011; 108(3):693-702.
- [37] S Ankanna, TNVKV Prasad, EK Elumalai, N Savithramma, *Dig J Nanomater Bios.* 2010; 5(2): 369-37.
- [38] B Harekrishna, KB Dipak, P Gobinda, S Priyanka, PD Sankar, P Santanu, M Ajay, *Colloid surface A* 2009; 348(3):212-216.
- [39] C. Krishnaraj, EG Jagan, R Ramchandran, SM Abirami, *process biochem* 2011
- [40] P. Rauwel, S. Kuunal, S. Ferdov and E. Rauwel, *Advances in Materials Science and Engineering*, 2015.
- [41] S. Ahmed, M. Ahmad, B.L. Swami, S. Ikram, *Journal of Advanced Research*, Vol 7, Issue 1, 2016, p.17-28.
- [42] V Saklani, Suman, V.K. Jain VK, *Microbial Synthesis of Silver Nanoparticles: A Review. J Biotechnol Biomaterial*, 2012.
- [43] E. Araujo, N.J. Andrade, L.H. Da Silva, P.C. Bernardes and A.V. Teixeira (2011), *Journal of Food Production*. 74 (4): 701-705.
- [44] C. M. D'Almeida and B. J. Roth, *Medical Applications of Nanoparticles*, Department of Physics, Oakland University, Rochester, Michigan.
- [45] S. Galdiero, A. Falanga, M. Vitiello, M. Cantisani, V. Marra and M. Galdiero, *Molecules* 2011, 16, 8894-8918.
- [46] Q.H. Tran, V.Q. Nguyen and A.T. Le, *Advances in Natural Sciences: Nanoscience and Nanotechnology*, 2013, Volume 4, Number 3.
- [47] S. Majeed, M.S. Abdullah, A. Nanda and M.T. Ansari, *Journal of Taibah University for Science*, 2016, Volume 10, Issue 4, p.614-620.
- [48] A. M. Shawkey, M. A. Rabeh, A. K. Abdulall3 and A. O. Abdellatif, *Advances in Life Science and Technology*, Vol 13, 2013.